

*10.537 Nanomaterial Characterization I*

# Chromatographic Methods

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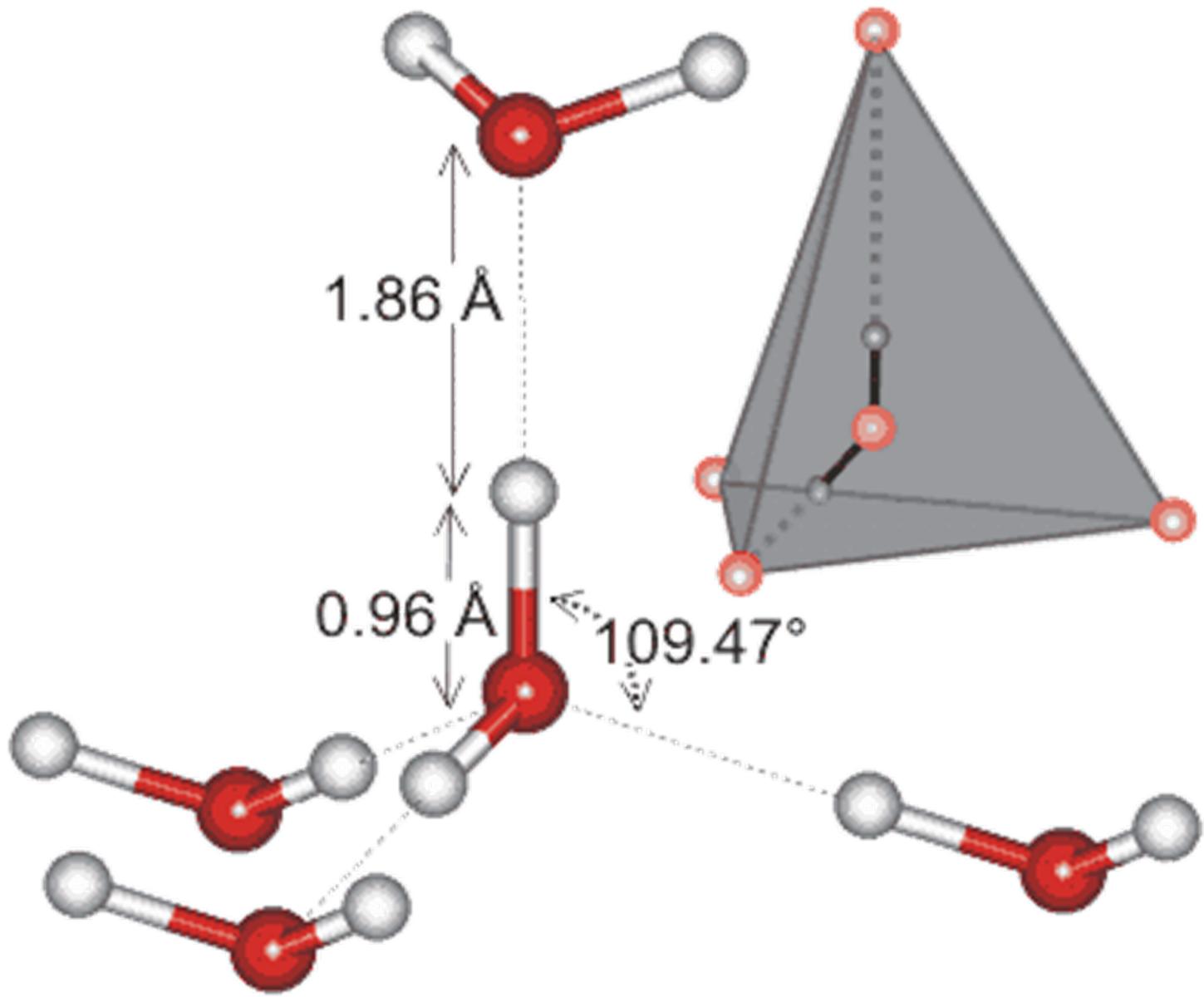
# Definitions

- **Analytical Chemistry** – qualitative and quantitative measurement of chemical species
- **Chromatography** – methods for separation and analysis of complex mixtures
- **HPLC** – High Performance Liquid Chromatography
- **GC** – Gas Chromatography

# Chromatographic Methods only work for dissolved molecular species

They are not directly applicable to:

- Nanoparticles
- Carbon Nanotubes
- Particulate or colloidal species including:
  - Graphene
  - Elemental nanoparticles
  - Etc.



# Introduction to Chromatography

- Chromatography is a separation technique
- HPLC & GC are our primary focus
- Also discuss low pressure column chromatography, not TLC (thin layer) or SFC
- All chromatographic techniques have
  - Stationary phase – solid or viscous liquid phase typically in a column
  - Mobile phase – moves sample in contact with stationary phase

Partitioning = type of equilibrium where the analyte divides itself between two phases

For liquid-liquid extraction – two liquids

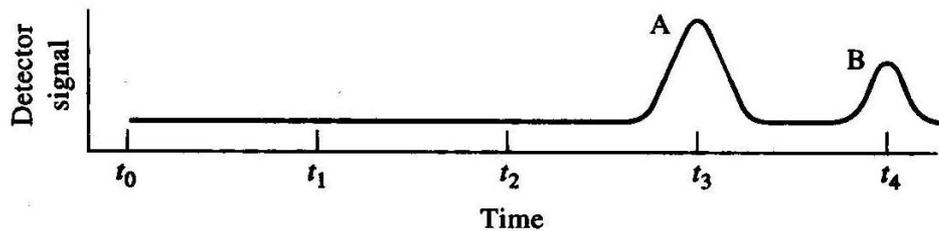
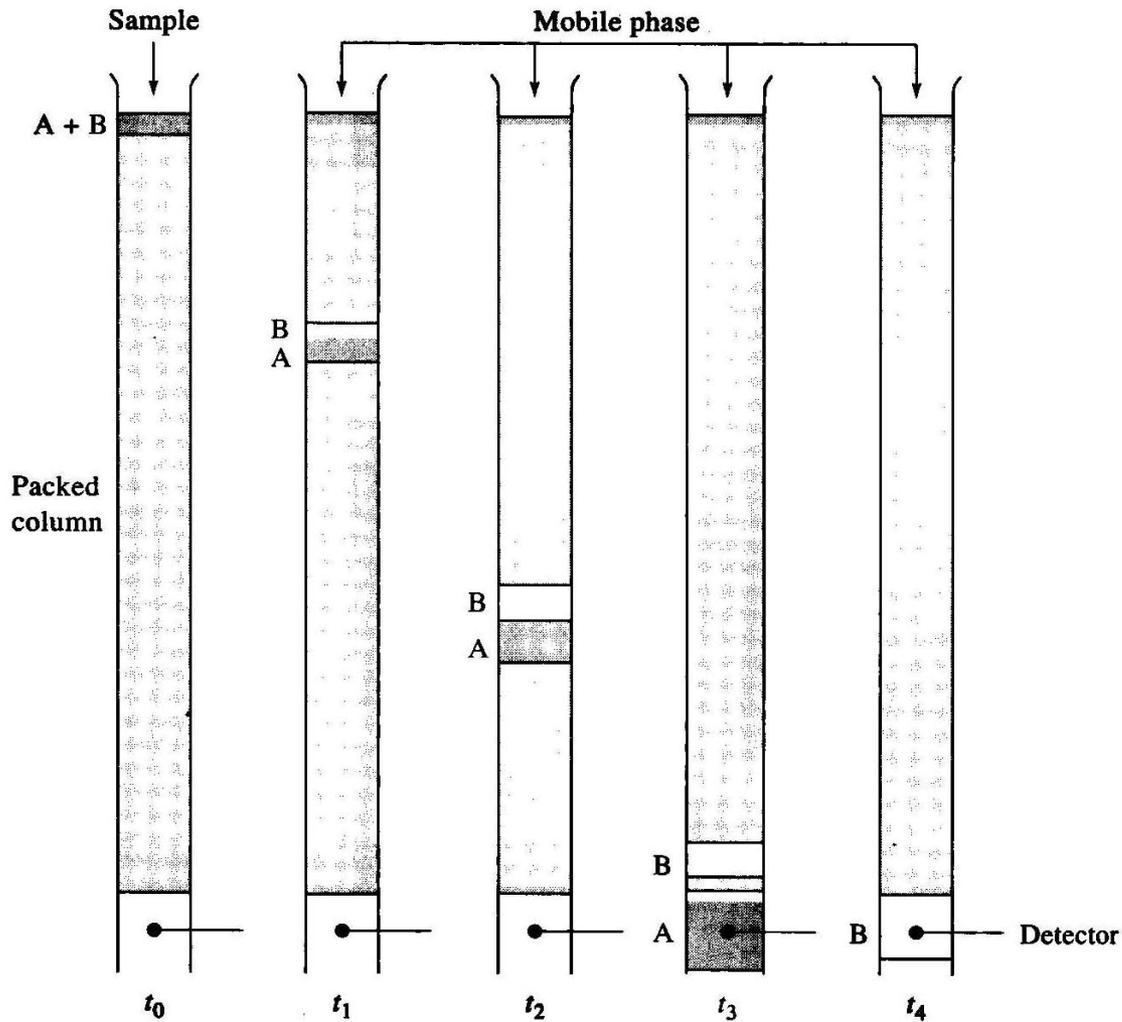
For chromatography – mobile vs. stationary phases

Define a partition ratio  $K$  (or distribution constant)

$$K = \frac{C_s}{C_M}$$

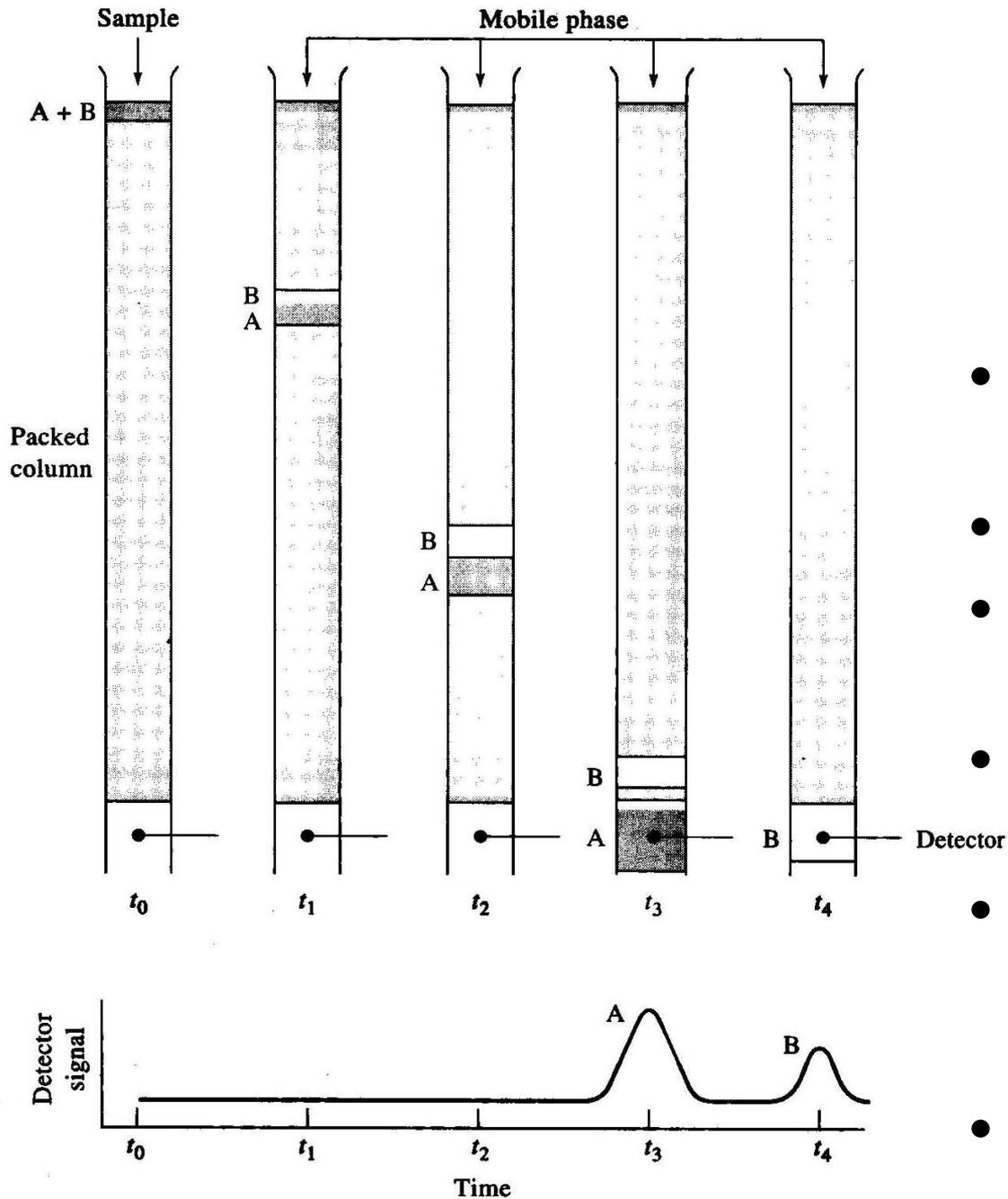
where  $C_s$  &  $C_M$  are concentrations of analyte in stationary & mobile phases <sub>6</sub>

- Prefer if  $K$  is constant over conc. range
- If not constant we can work in a narrow range where it is constant
- This is linear chromatography
- In linear chromatography a constant flow rate of mobile phase moves through column
- Elution = process by which analyte is flushed through the column by mobile phase (which could be a liquid or a gas)

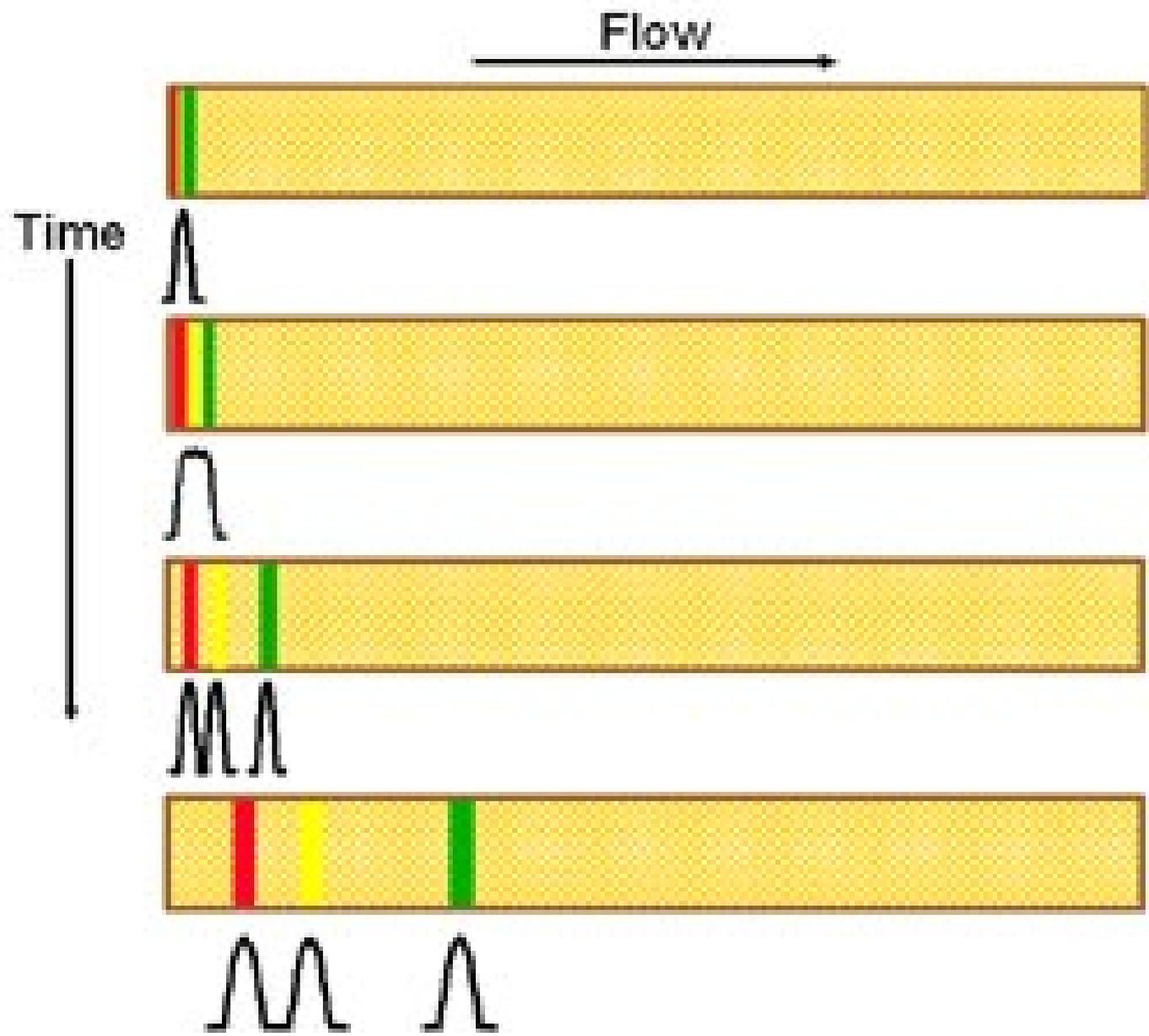


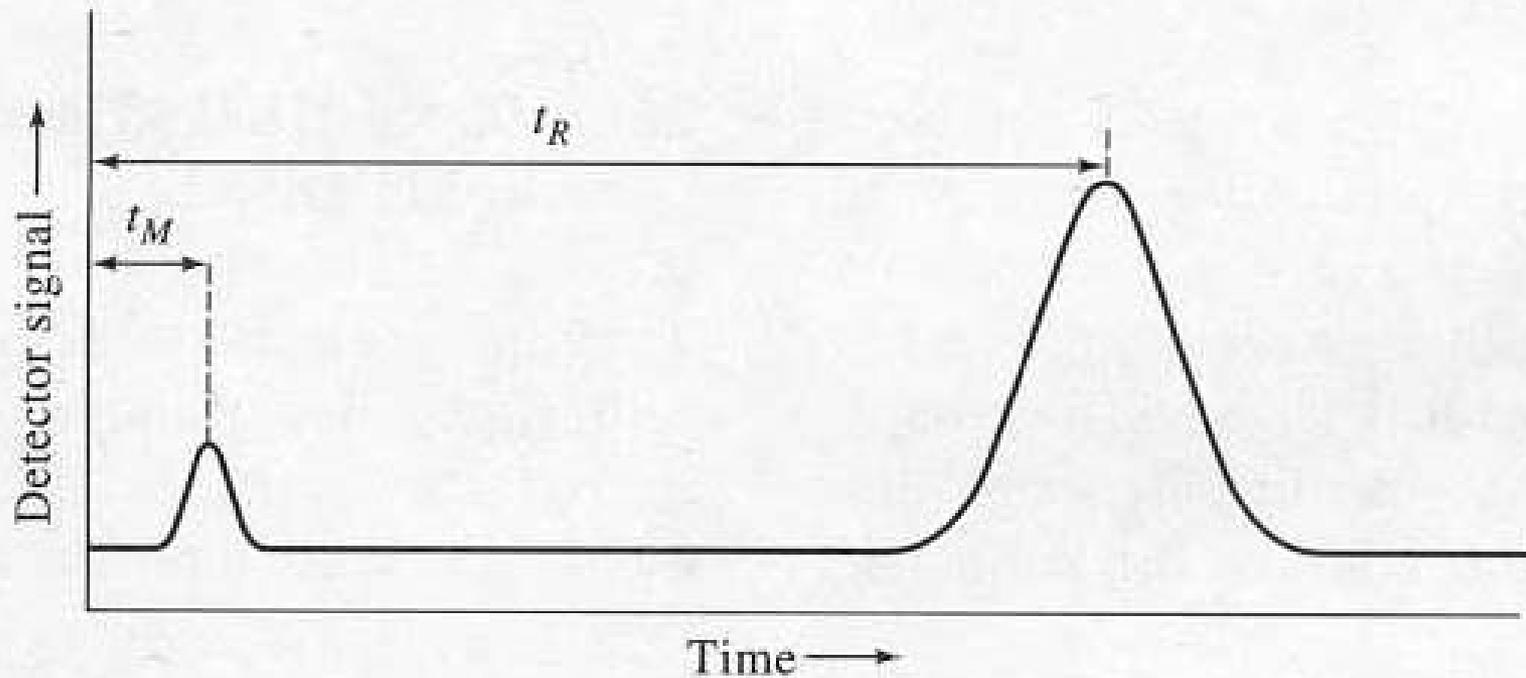
## Overview of chromatographic process – packed column

- Inject at  $t_0$
- Separate  $t_1$  to  $t_3$
- Detect at  $t_4$
- ← Resulting chromatogram



- A & B retained by column differently
- B has higher K
- B takes longer to elute from column
- Detector sees A first then B
- Peak heights & peak areas are proportional to conc.
- Band broadening 9





**Figure 26-4** A typical chromatogram for a two-component mixture. The small peak on the left represents a species that is not retained on the column and so reaches the detector almost immediately after elution is started. Thus its retention time  $t_M$  is approximately equal to the time required for a molecule of the mobile phase to pass through the column.

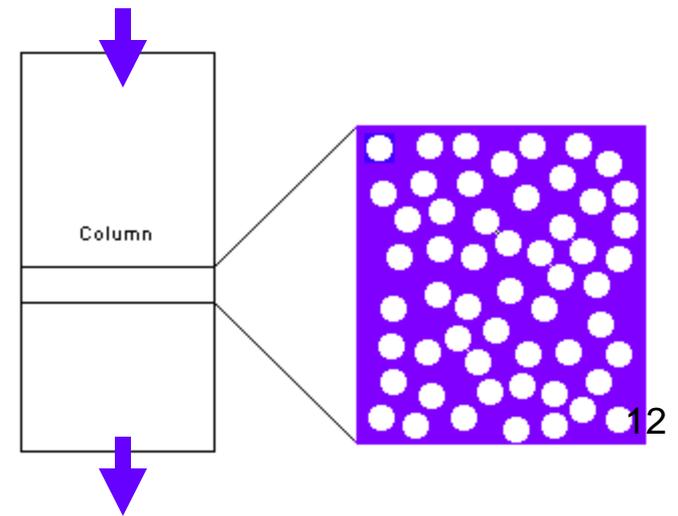
$t_M$  = time for unretained molecule to reach detector or dead time

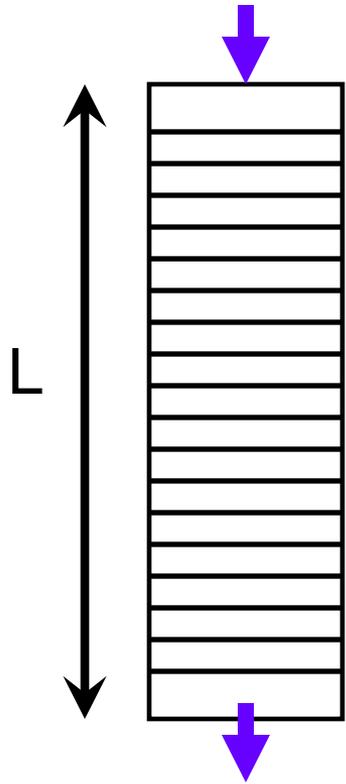
$t_R$  = retention time, time for retained species to reach detector<sub>1</sub>

# Chromatographic Plate Theory vs. Rate Theory

- Plate theory based in liquid-liquid extraction (successive extractions)
- $K = C_{\text{org}}/C_{\text{water}}$
- Chromatographic column can be thought of in the same way (only continuous process)
- $K = C_s/C_M$

- Stationary phase bead
- Mobile phase (liquid)





$$L = NH$$

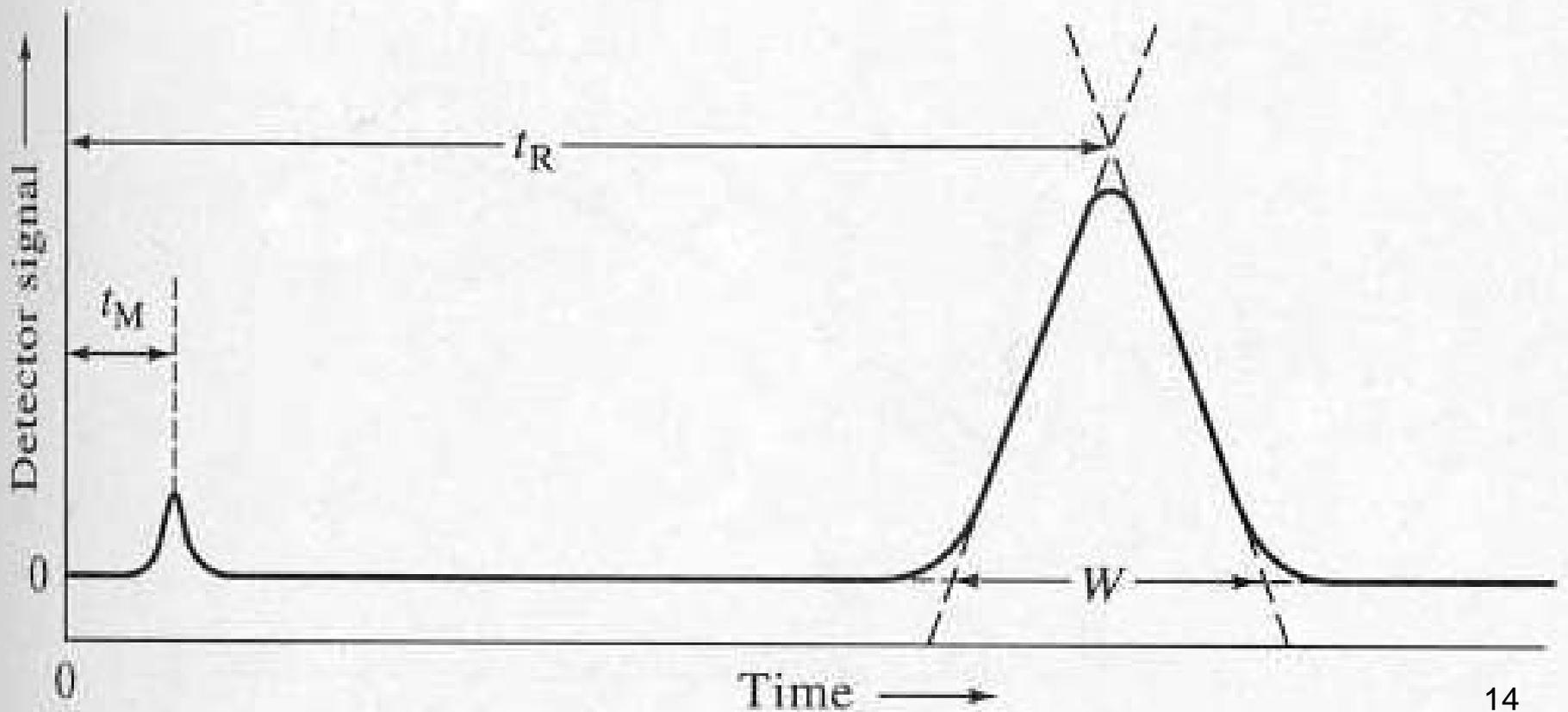
or

$$N = L/H$$

- Divide chromatographic column up into steps or segments called theoretical plates
- The theoretical concept is that these theoretical plates are equilibrium units for  $K = C_s/C_M$
- The more theoretical plates a column has, the more efficient it is
- If column length =  $L$  &  $N$  = number of plates, then  $H$  = height equivalent to theoretical plate

# Gaussian distribution (bell curve)

$$W = 4\sigma$$



Can derive

N = number of plates

$$N = 16 (t_R/W_b)^2$$

$W_b$  = base width

$$N = 16 (t_R/4\sigma)^2 = (t_R/\sigma)^2$$

$$N = 5.54 (t_R/W_{1/2})^2$$

$W_{1/2}$  = width at  
half height

Column manufacturers use N

to characterize column – N varies widely

# Shortcomings of Plate Theory

- Assumes  $K$  is independent of concentration
- Assumes equilibration is rapid relative to velocity of mobile phase – not true, in reality solute may pass a plate without entering
- Assumes no longitudinal diffusion (= non ideal effect that causes band broadening)
- Does not address several factors caused by mobile phase velocity (fast or slow) Rate Theory
- Assumes discrete units or plates for equilibrium rather than a semi continuous process through the column

# Rate Theory of Chromatography

$$H = H_L + H_S + H_M + H_{SM}$$

$H$  = height equivalent to theoretical plate (as in Plate Theory)

$H_L$  = contribution due to longitudinal diffusion

$H_S$  = stationary phase mass transfer contribution

$H_M$  = diffusion associated with mobile phase effects

$H_{SM}$  = diffusion into or mass transfer across a stagnant layer of mobile phase (neglect)

$$H = B/\mu + C\mu + A$$

van Deemter Equation  $A$ ,  $B$  &  $C$  are coefficients,  $\mu$  = velocity

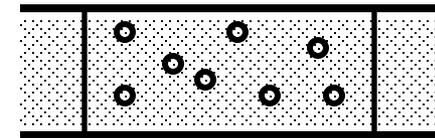
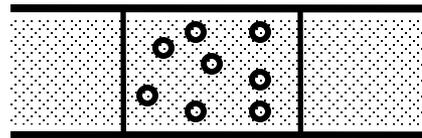
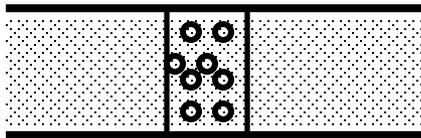
# 1) Longitudinal Diffusion

$$H_L = (B/\mu)$$

$t = 0$

$0 < t < t_R$

$t_R$



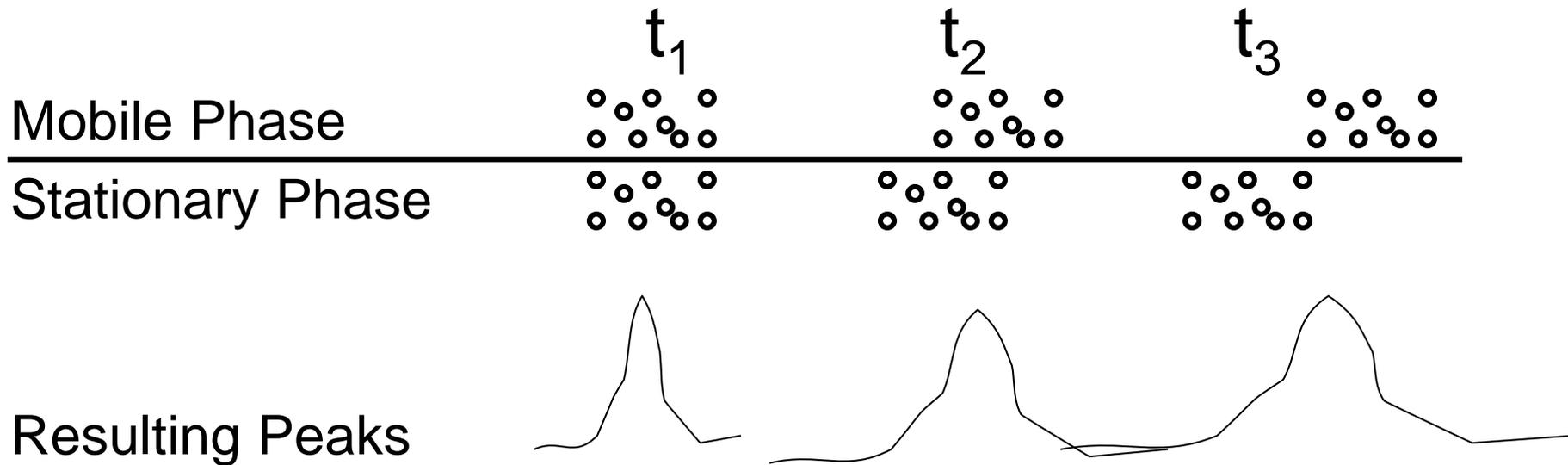
$$\sigma_L^2 = 0$$

$$\sigma_L^2 = 2 D_M t_M$$

Variance due to longitudinal diffusion = 0 at start

Variance increases with time & diffusion coefficient D

## 2) Mass transfer in & out of stationary phase

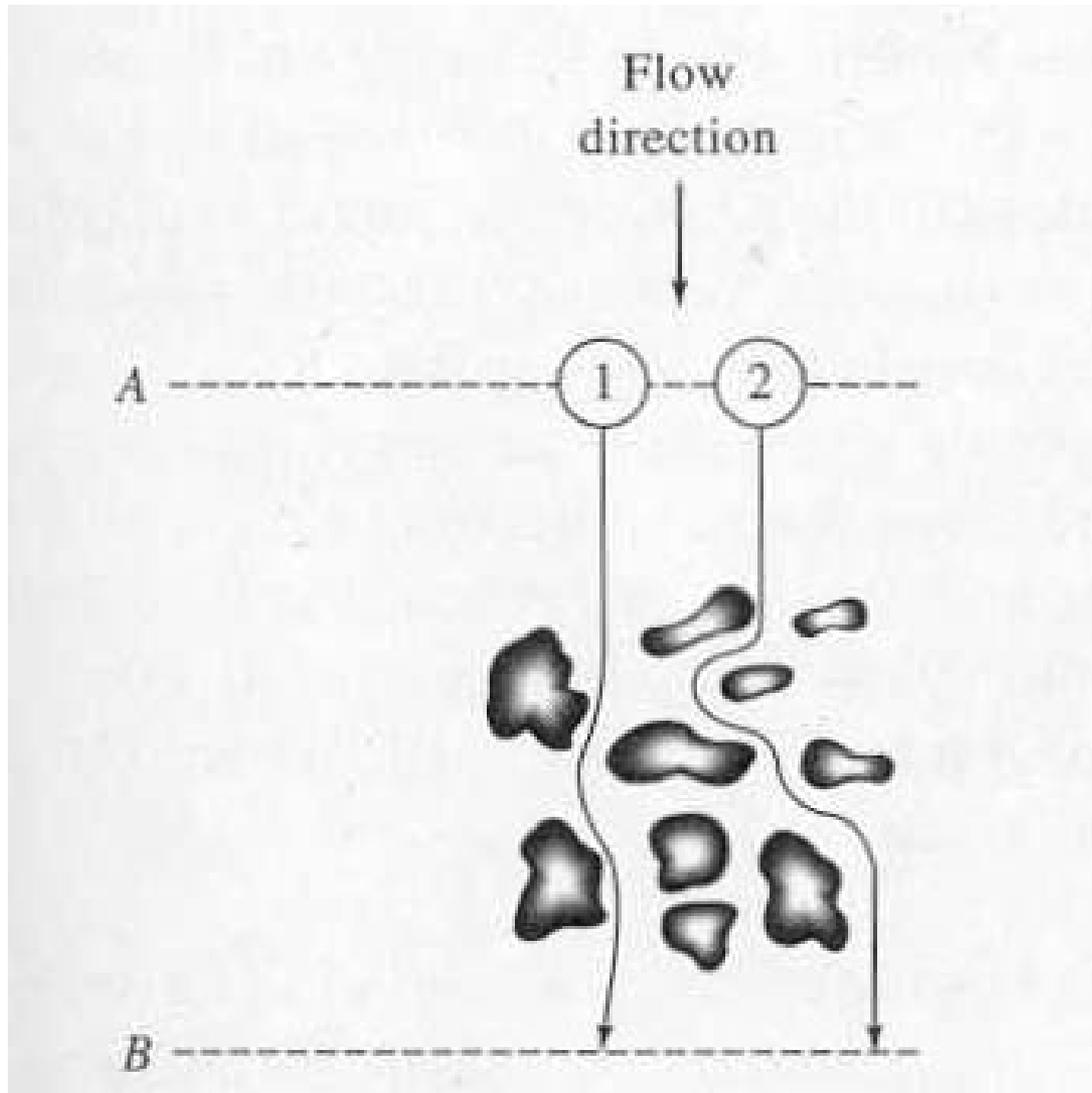


Broadening of peaks is a function of mobile phase velocity  
(moving molecules faster than those in stationary phase)

Not the same as longitudinal diffusion  $H_S = C\mu$

In Plate Theory condition at  $t_1$  assumed to hold throughout<sup>19</sup>

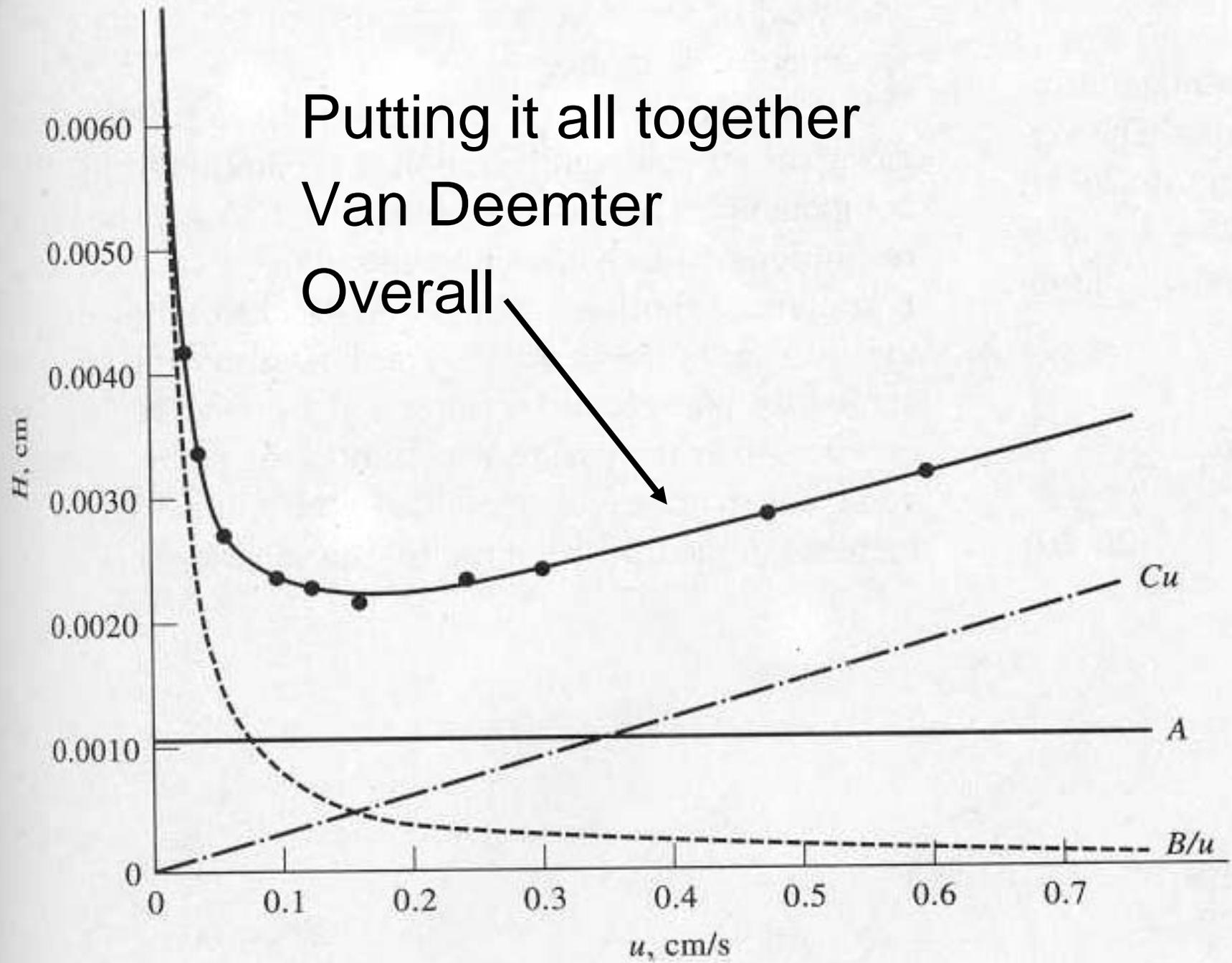
### 3) Uneven Flow or Eddy Diffusion



Path 1 is shorter than path 2

$$H_M = A$$

Putting it all together  
Van Deemter  
Overall

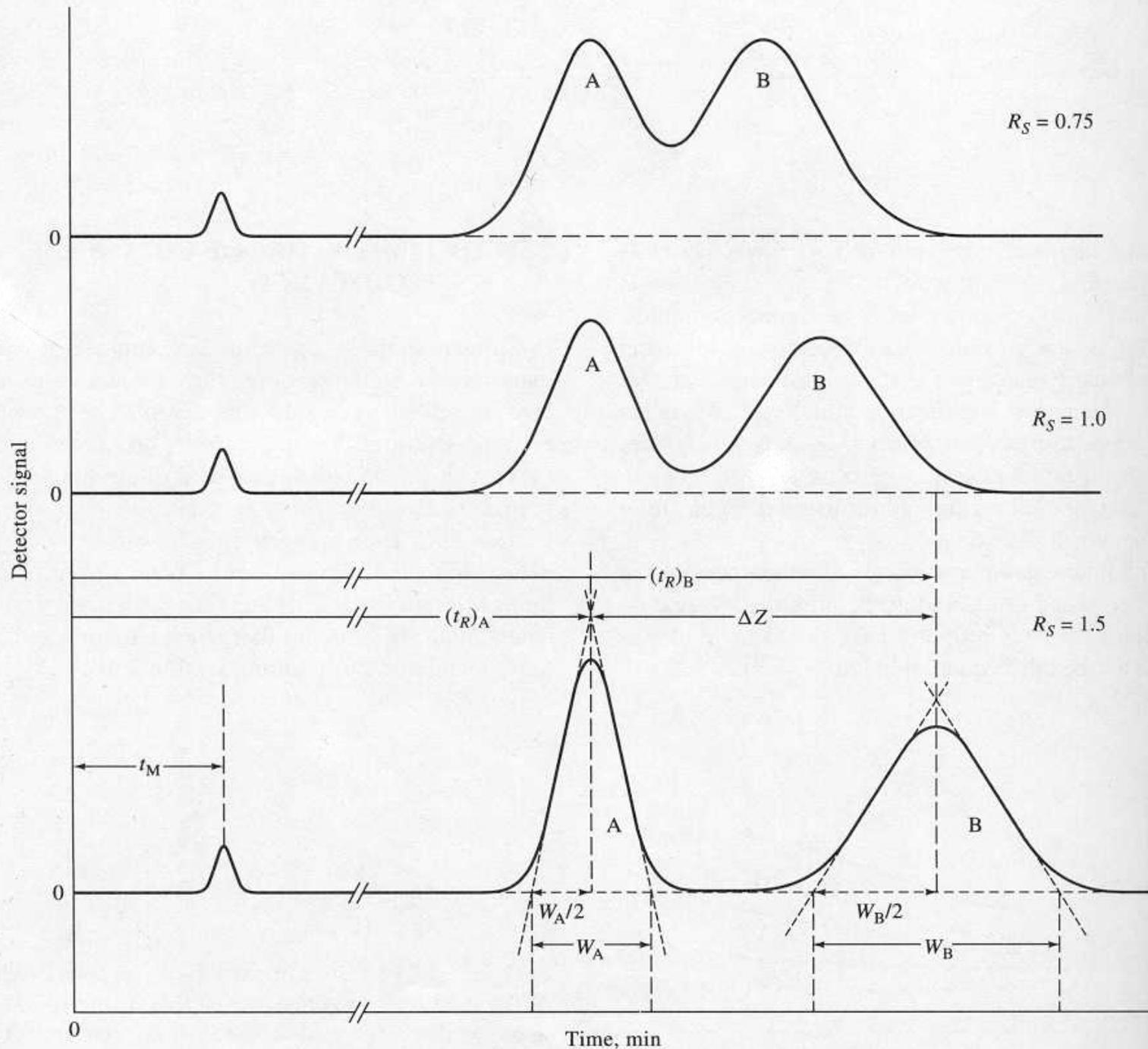


Optimizing Column Performance – seldom operate at optimum → too slow

Normally want to get required separation in shortest time, this may be at  $2X \mu_{opt}$

Can optimize a separation by varying experimental conditions, usually goals are

- 1) reduce band broadening (zone)
- 2) alter relative migration rates of components (allowing better separation of two components)



This brings us to Resolution ( $R_S$ ) = Measure of columns ability to separate 2 analytes

Note  $\Delta Z$  = spread of peaks &  $W$  or  $W/2$  = peak width

$$R_S = \frac{\Delta Z}{W_A/2 + W_B/2} = \frac{2 \Delta Z}{W_A + W_B} = \frac{2[(t_R)_B - (t_R)_A]}{W_A + W_B}$$

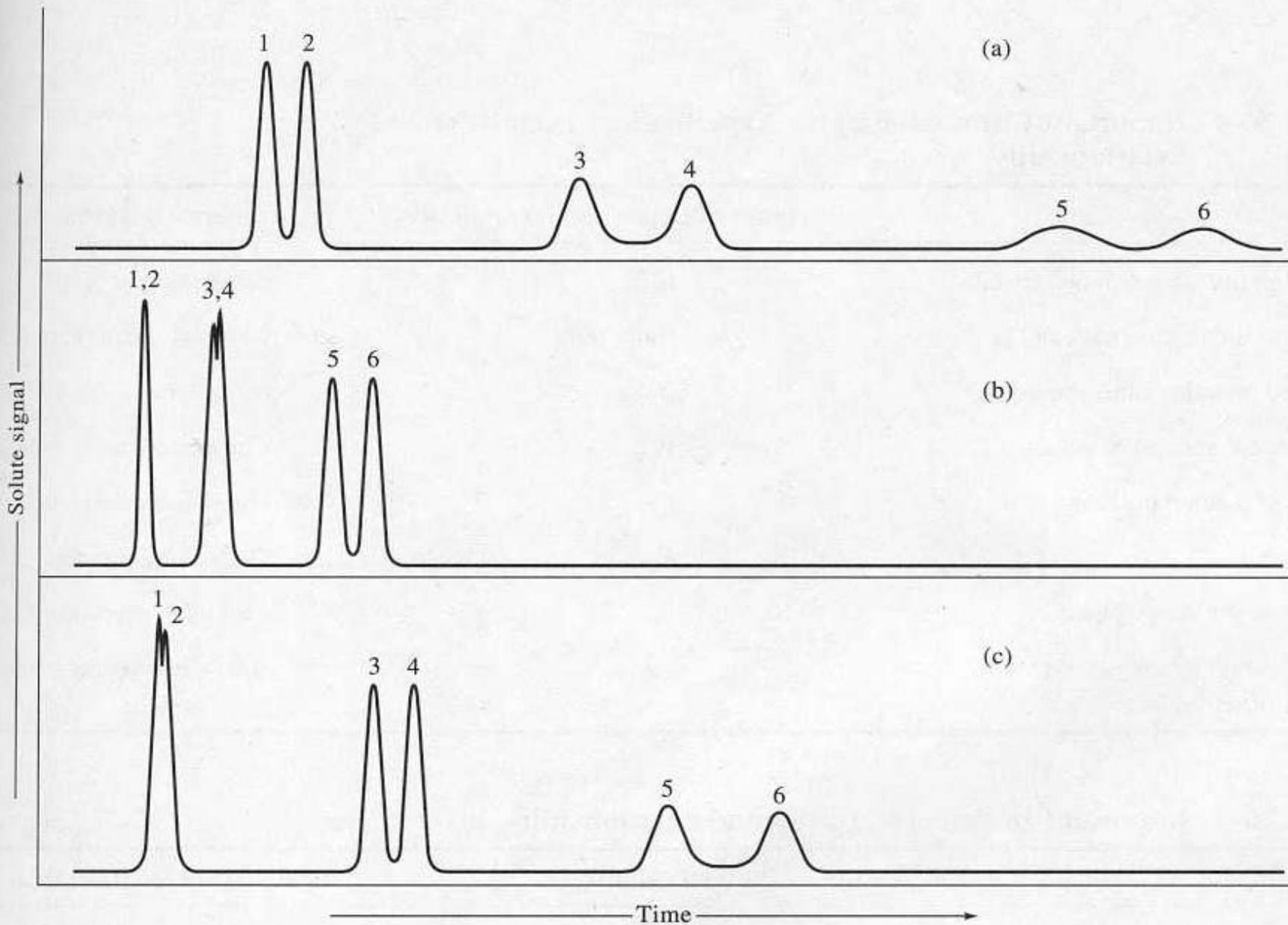
If  $R_S = 1.0$  then  $\Delta Z = W_A/2 + W_B/2$

and peaks touch with about 4% overlap

This is too big an error to tolerate

If  $R_S = 1.5$  then about 0.3% overlap

Can lengthen column to improve resolution by increasing  $N \rightarrow$  this also increases time for analysis



**Figure 26-14** Illustration of the general elution problem in chromatography.

Commonly found problem in chromatography

General Elution Problem

Solution – change conditions during chromatographic run so that  $k'$  changes

Start with conditions for chromatogram (a), after 1 & 2 elute

Change to conditions for chromatogram (c), after 3 & 4 elute

Change to conditions for chromatogram (b) to get 5 & 6

To Solve the General Elution Problem:

In GC do temperature programming

In HPLC do solvent programming (a.k.a.  
gradient elution)

# Gas Chromatography

- Principles
- Instrumentation
- Detectors
- Columns and Stationary Phases
- Applications

**Basic Principle of GC** – sample vaporized by injection into a heated system, eluted through a column by inert gaseous mobile phase and detected

**Three types (or modes)**

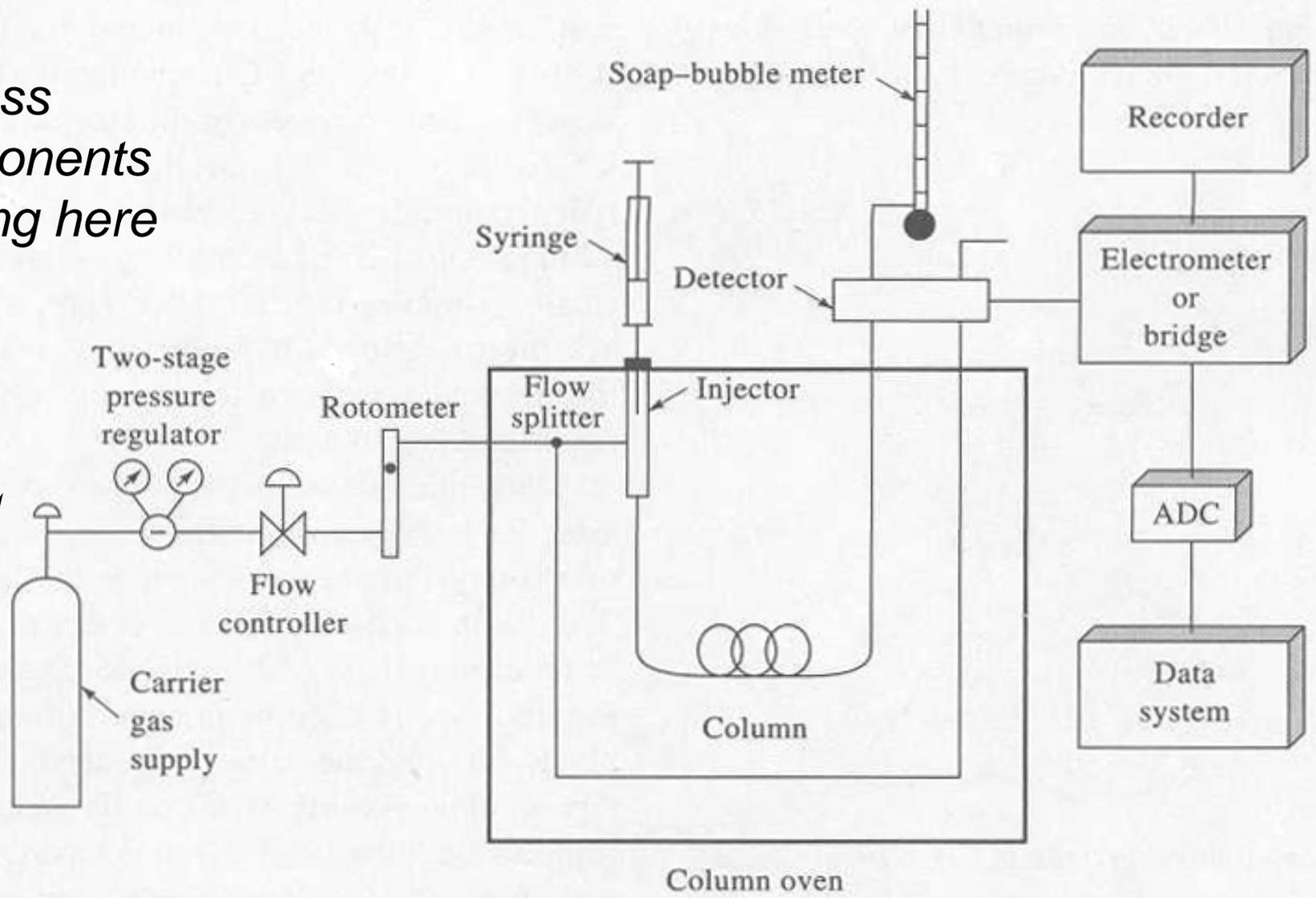
gas – solid chromatography ← early

gas – liquid “ ← important

gas – bonded phase “ ← relatively new

An estimated 200,000 GC in use worldwide <sub>29</sub>

*Discuss components starting here*



**Figure 27-1** Schematic of a gas chromatograph.

Carrier gases (mobile phase) – must be chemically inert He, Ar, N<sub>2</sub>, CO<sub>2</sub> even H<sub>2</sub> and mixtures 95/5 N<sub>2</sub>/CH<sub>4</sub>

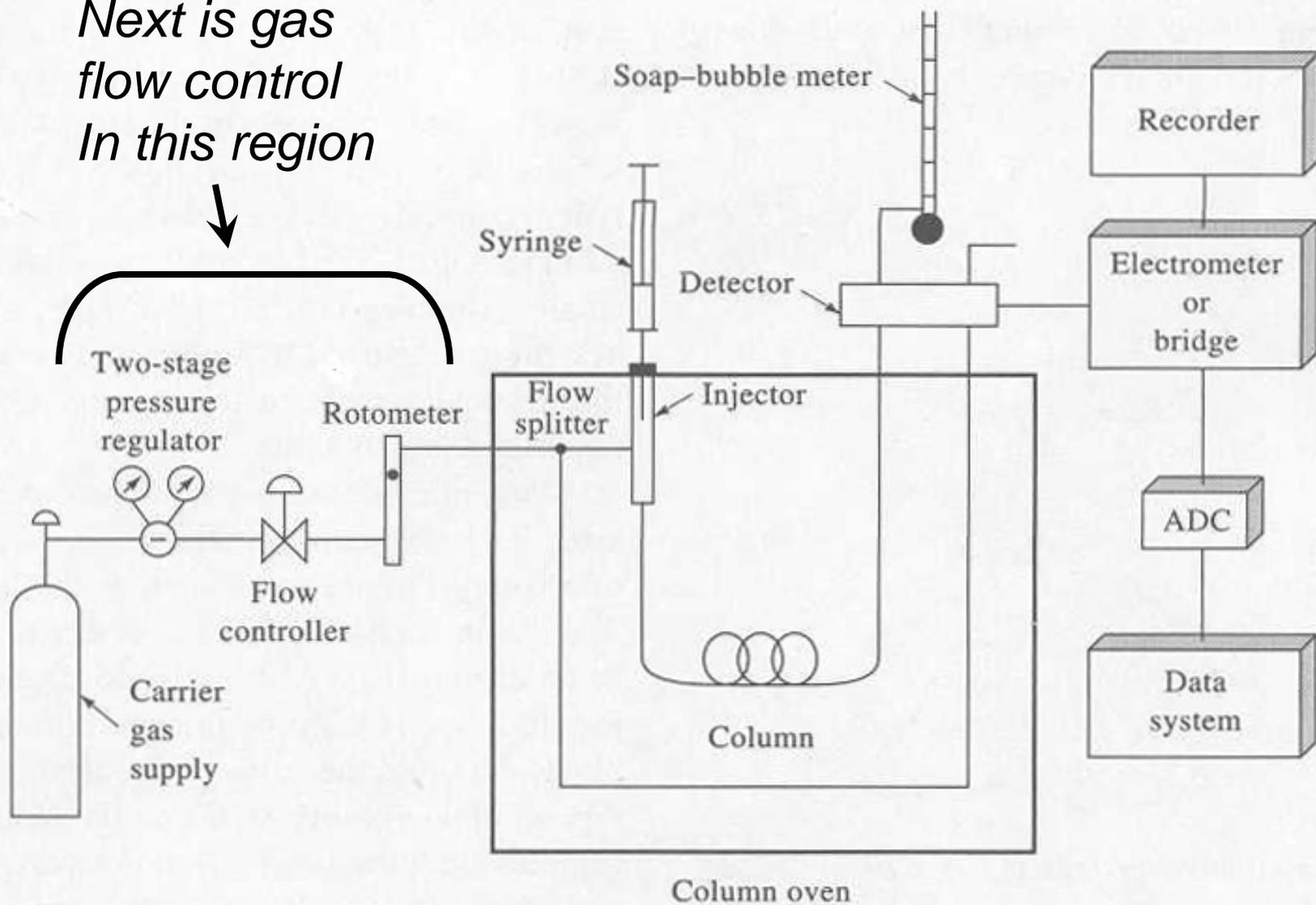
Often detector dictates choice of carrier gas

In GC sample doesn't really interact with carrier gas (unlike HPLC), temp controls partitioning

Often necessary to purify cylinder gas with a trap, scrubber or cartridge of molecular sieves (or buy high purity gas) O<sub>2</sub> ppm Hc

The move today is away from gas cylinders toward gas generators (extract pure carrier gas from air)

*Next is gas  
flow control  
In this region*

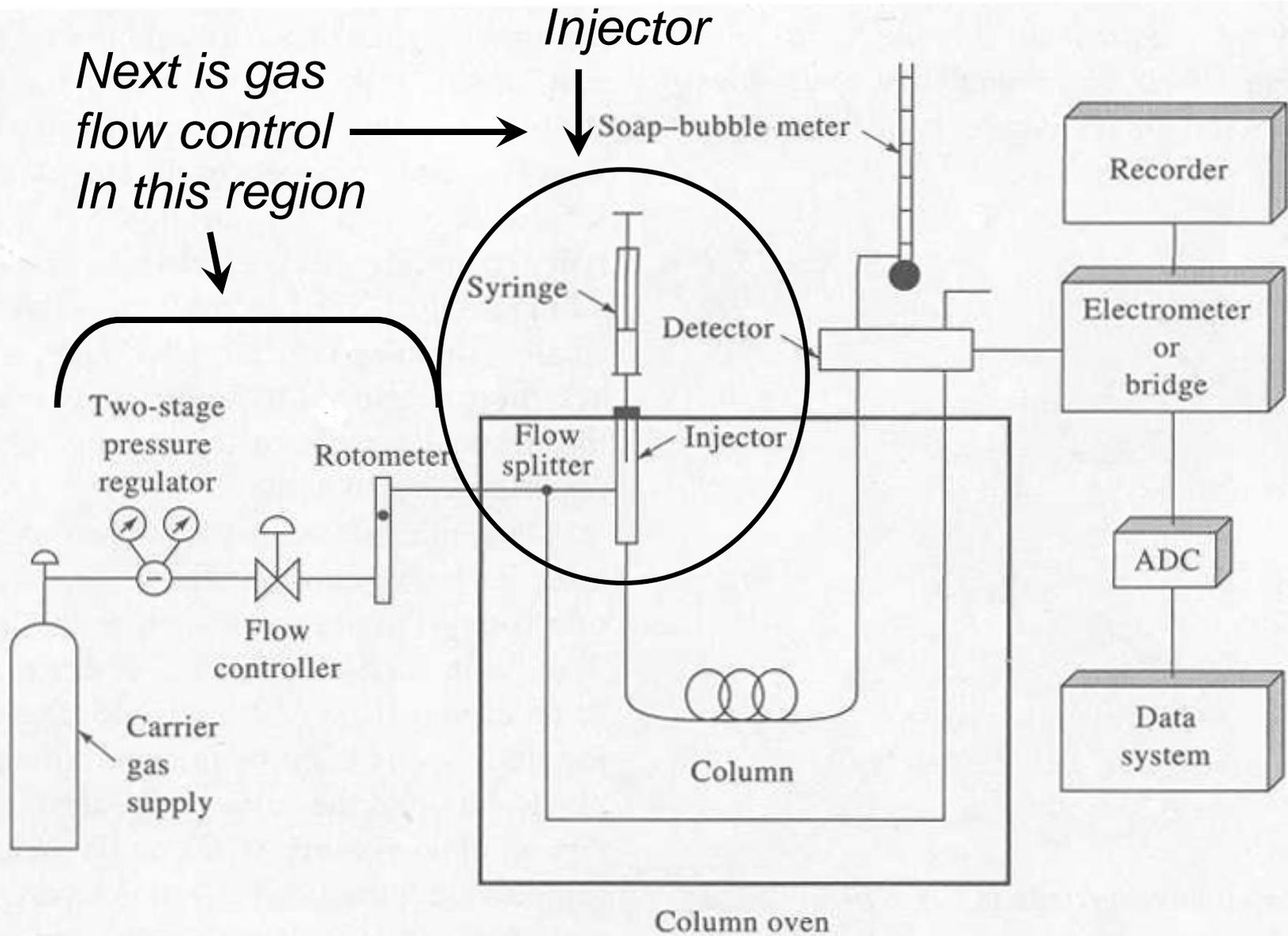


**Figure 27-1** Schematic of a gas chromatograph.

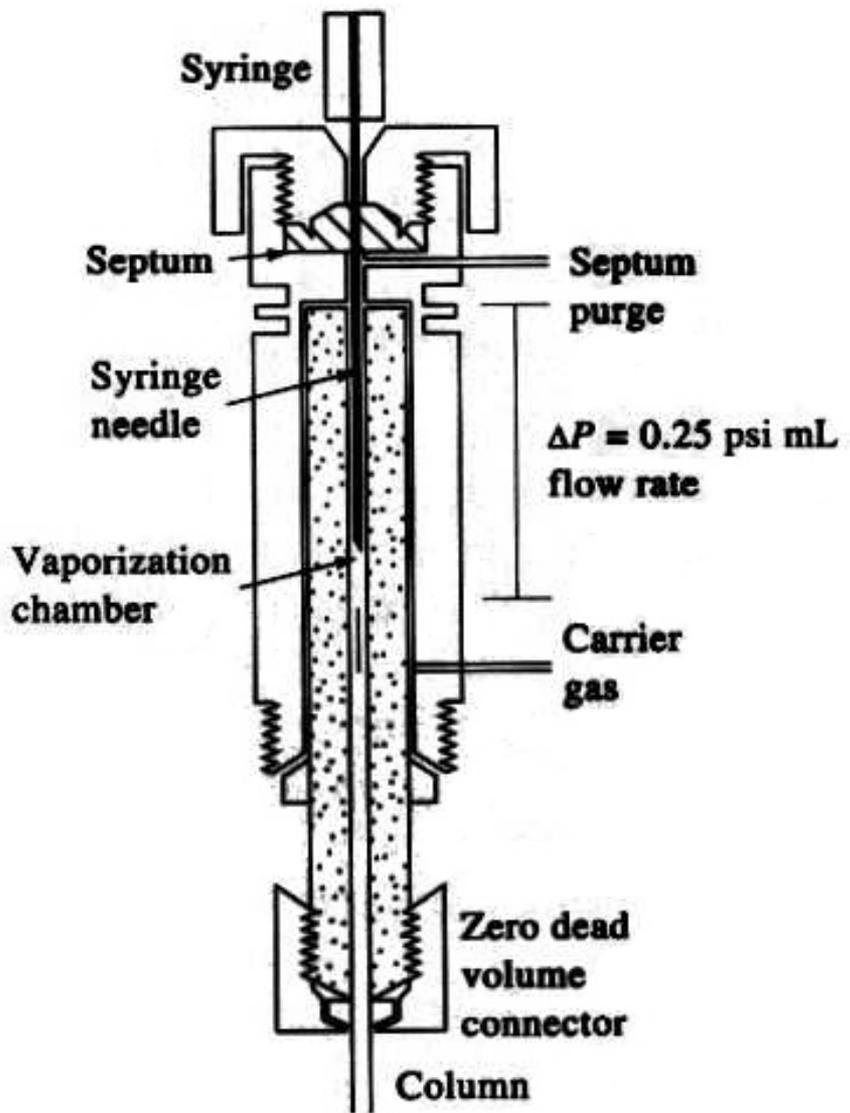
Flow control – 10 to 50 psi with regulator

Regulators vary in quality, material & control,  
typically use a 2 stage regulator with the  
best material being stainless steel

Ultimately flow rate is checked by a soap  
bubble meter for accurate flow



**Figure 27-1** Schematic of a gas chromatograph.



**Figure 27-3** Cross-sectional view of a microflash vaporizer direct injector.

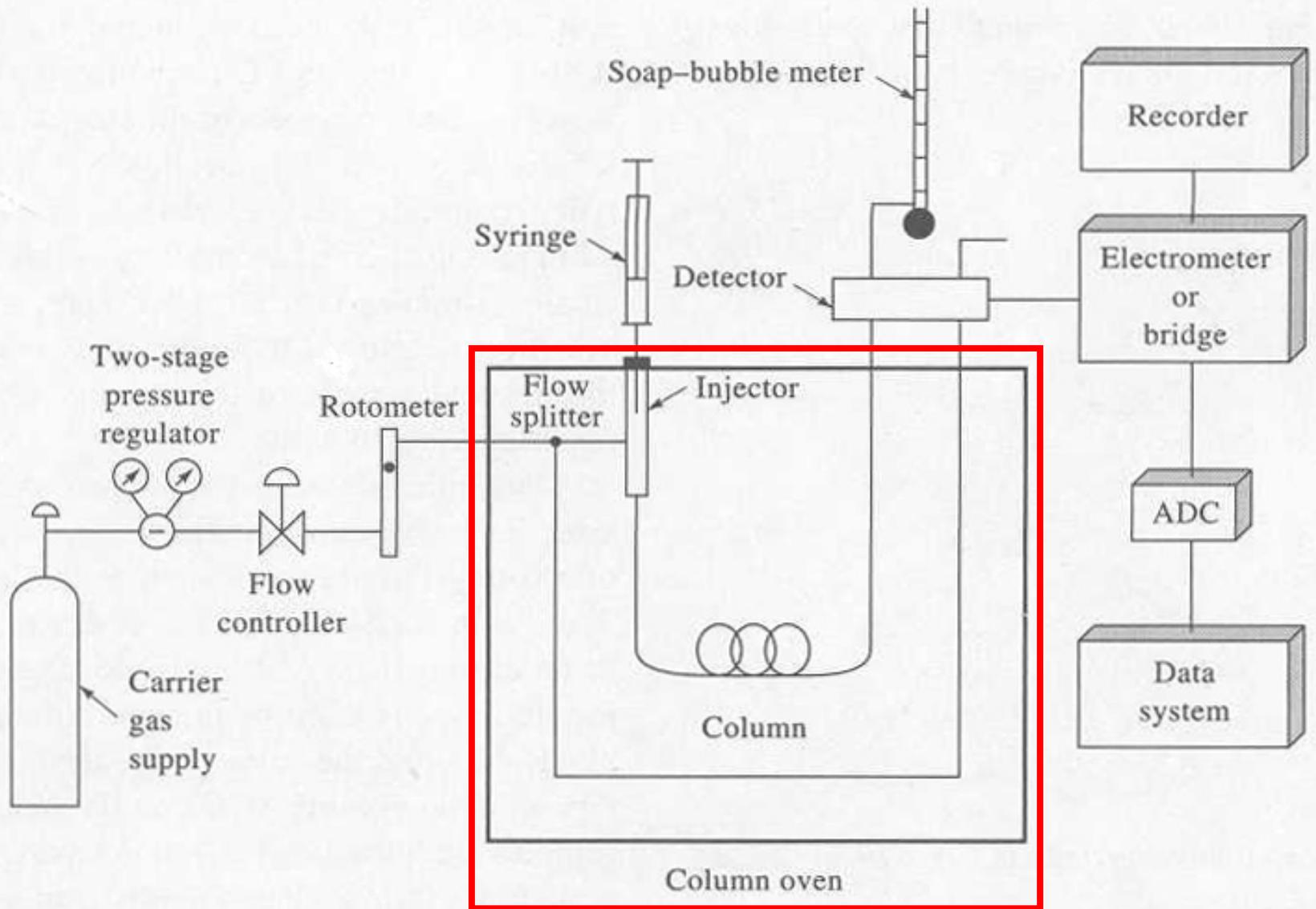
Injector – use micro syringe 99.9 % of the time injecting 1 to 20  $\mu\text{L}$ , rapidly shoot in plug of sample

Old GCs had separate injection area

Today use on-column & microflash vaporizers – all have septum of synthetic rubber which is punctured by syringe

Injector usually 50  $^{\circ}\text{C}$  hotter than boiling point of sample – also hotter than column

Can use rotary injector valve (as for HPLC)



**Figure 27-1** Schematic of a gas chromatograph.

Column housed in Column Oven to maintain temperature

Types – packed, open tubular, capillary  
oldest ----- newest

Capillary columns will take over completely

Packed – tube (steel, glass, **fused silica**, Teflon) packed with material

Open Tubular – coated on walls

Capillary – coated on walls, long & narrow

Length range – 2 to 50 m (typically 30 m)

# Detectors – dozens of detectors available

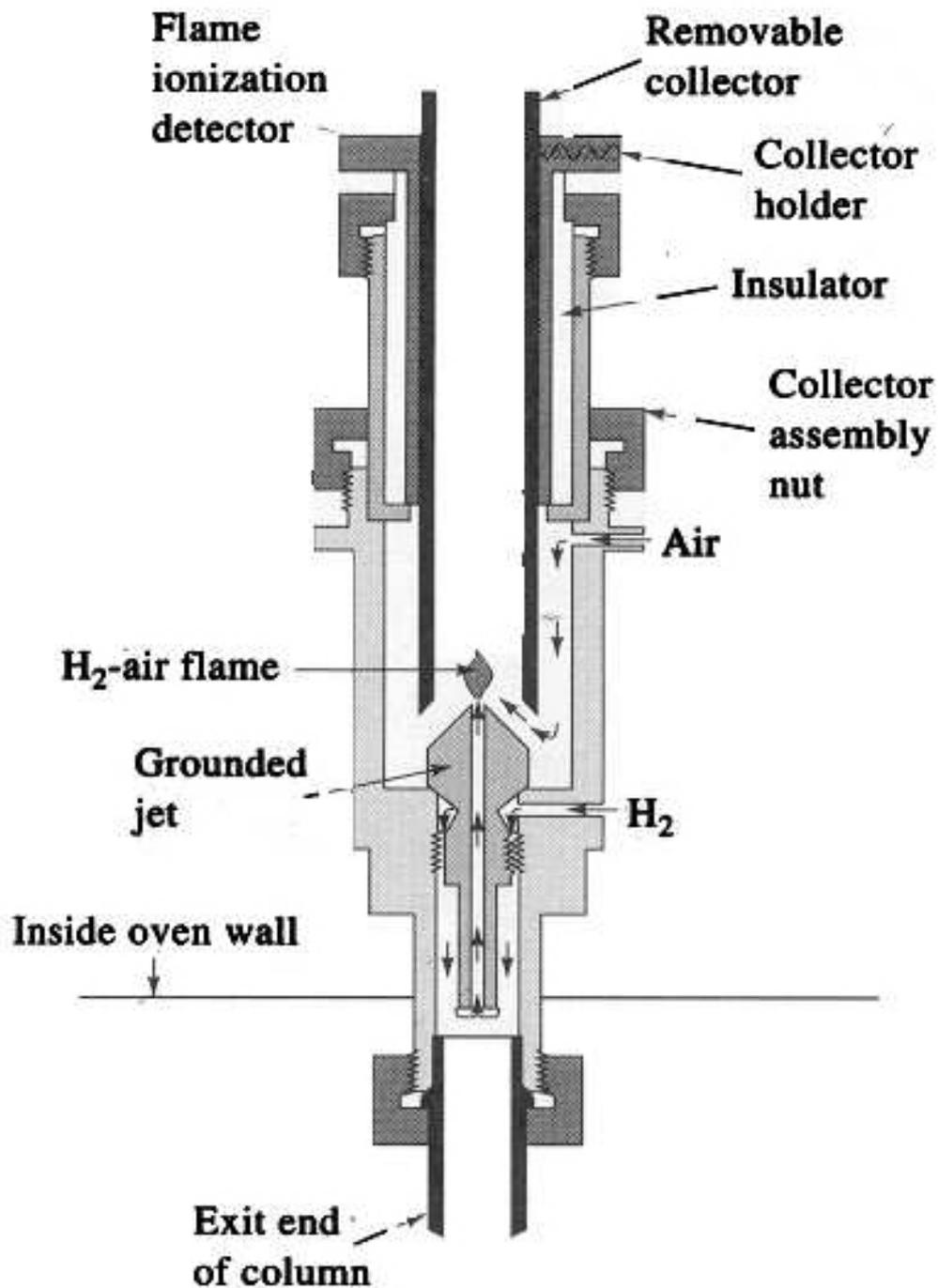
## Characteristics of an ideal detector:

- 1) Adequate sensitivity for desired analysis  
(typical  $10^{-8}$  to  $10^{-15}$  g analyte/sec)
- 2) Stable – background constant with time
- 3) Reproducible – good precision
- 4) Linear response over several orders of magnitude
- 5) Temperature range – room temp - 400 °C

## Characteristics of ideal detector: (continued)

- 6) Rapid response time
- 7) Independent of flow rate
- 8) Reliable
- 9) Easy to Use – inexperienced operators
- 10) Either selective or universal response
- 11) Nondestructive

No detector exhibits all these characteristics



## Flame Ionization Detector (FID)

- one of most widely used GC detectors
- good sensitivity to almost all organic compounds

# FID Basics

- column effluent mixed with air and burned in H<sub>2</sub> flame producing ions & electrons that conduct electricity
- a few hundred volts applied between burner tip & a collector electrode above the flame producing currents on the order of 10<sup>-12</sup> amps
- amplify & measure
- signal approximately proportional to number of reduced carbon atoms in flame

## FID Basics (continued)

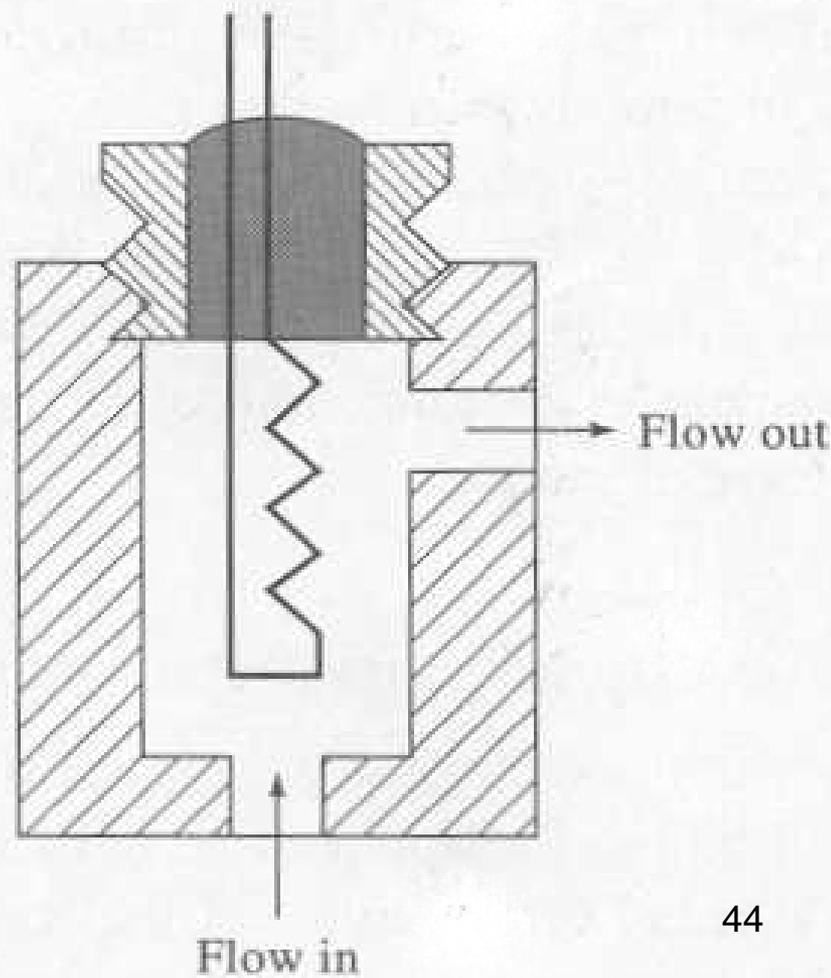
- mass sensitive rather than concentration
- insensitive to non combustible gases –  
 $\text{H}_2\text{O}$ ,  $\text{CO}_2$ ,  $\text{SO}_2$ ,  $\text{NO}_x$

## FID exhibits

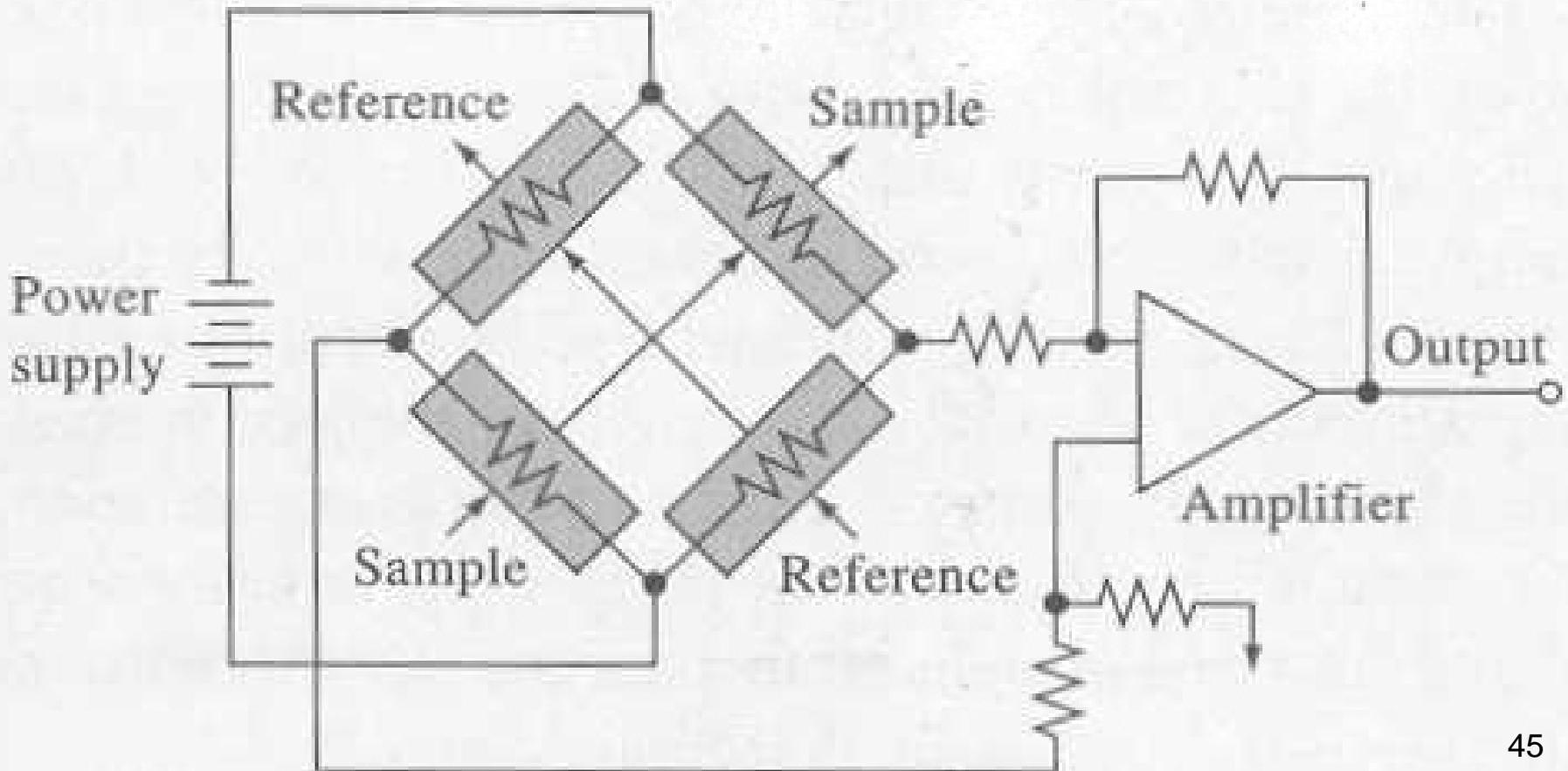
- High sensitivity (as low as  $10^{-13}$  g/s)
- Large linear response range ( $10^7$ )
- Easy to use
- Rugged
- **DESTRUCTIVE**

# Thermal Conductivity Detector (TCD)

- One of earliest GC detectors
- Not popular today
- Low sensitivity
- Several designs
- Use heated wire or semiconductor
- Resistance of wire changes with analyte vs carrier



# TCD uses bridge circuit with Sample & Reference Cells



# TCD

- New TCDs use pulsed current to increase sensitivity & reduce drift
- Thermal conductivity of He & H<sub>2</sub> are about 6 to 10 times greater than most organic compounds (must use these carrier gases)
- Other carrier gases (N<sub>2</sub>, Ar, etc) have thermal conductivities too close to organics

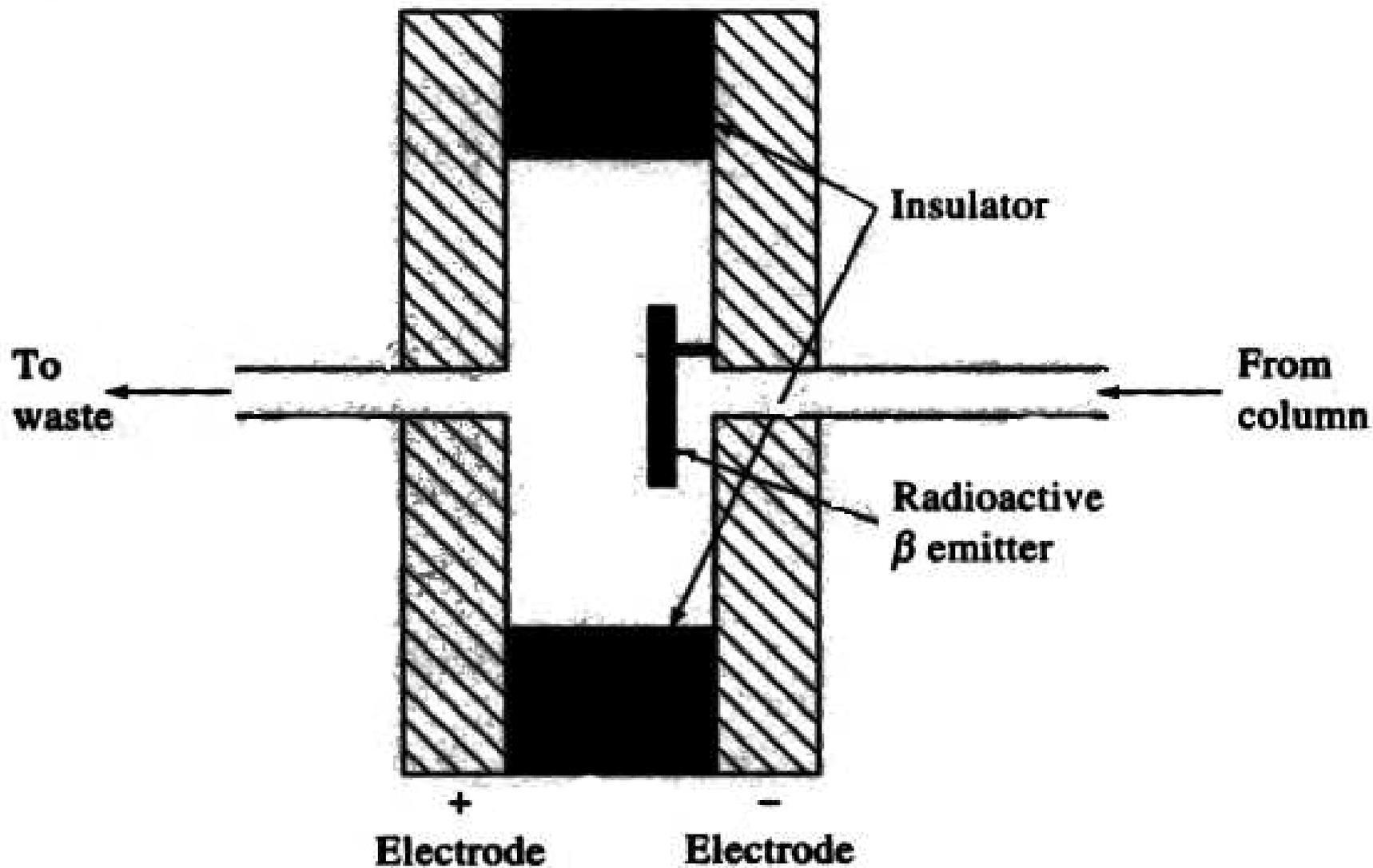
## Advantages of TCD

- Simple → Reliable & Easy to use
- Universal response (organic & inorganic)
- Large linear dynamic range  $10^5$
- Nondestructive, can use in tandem
- Older instruments have built-in TCD

## Disadvantages

- Low sensitivity
- Often can't use with capillary columns because amount of analyte is small

# ECD



**Figure 27-8** A schematic of an electron-capture detector.

# Electron Capture Detector

- Sample passes over  $\beta$  emitter (radioactive) like  $^{63}\text{Ni}$  foil or  $^3\text{H}_2$  adsorbed on Pt or Ti foil
- $\beta$  particles (i.e. electrons) hit carrier gas (usually  $\text{N}_2$ ) causing a burst of  $e^-$  to be released & measured by electrode = standing current or constant signal
- When analyte molecule that absorbs  $e^-$  passes through, current is reduced = signal
- Response is non-linear unless pulsed

# ECD Advantages

- Responds well to molecules with electronegative atoms like halogens (F, Cl, Br, I), peroxides, quinones, & nitro groups
- Insensitive to amines, alcohols, hydrocarbons
- Chlorinated pesticides are big application
- Highly sensitive
- Easy to use
- Pretty reliable, although foil can get coated
- Selective

## ECD Disadvantages

- Narrow linear range
- Radioactive
- Regular wipe test
- Bake out contaminants
- Some limits to applicability because highly selective

# Other Conventional Detectors

## Thermionic Detector (TID)

- Selective for N & P compounds
- 500 x more sensitive than FID for P
- 50 x more sensitive than FID for N
- Bad for C
- Design similar to FID with rubidium silicate bead at 180 V vs collector → get hot plasma 600 - 800 °C
- Produces large number of ions with N & P

# Flame Photometric Detector (FPD)

- Selective for P & S compounds
- Again sample goes through H<sub>2</sub>/air flame
- Observe optical emission of HPO at 510 nm & 526 nm & S<sub>2</sub> at 394 nm
- Use optical filters to isolate signal
- Can also measure halogens, N, some metals (e.g. Cr, Ge, Se)

# Photoionization Detector (PID)

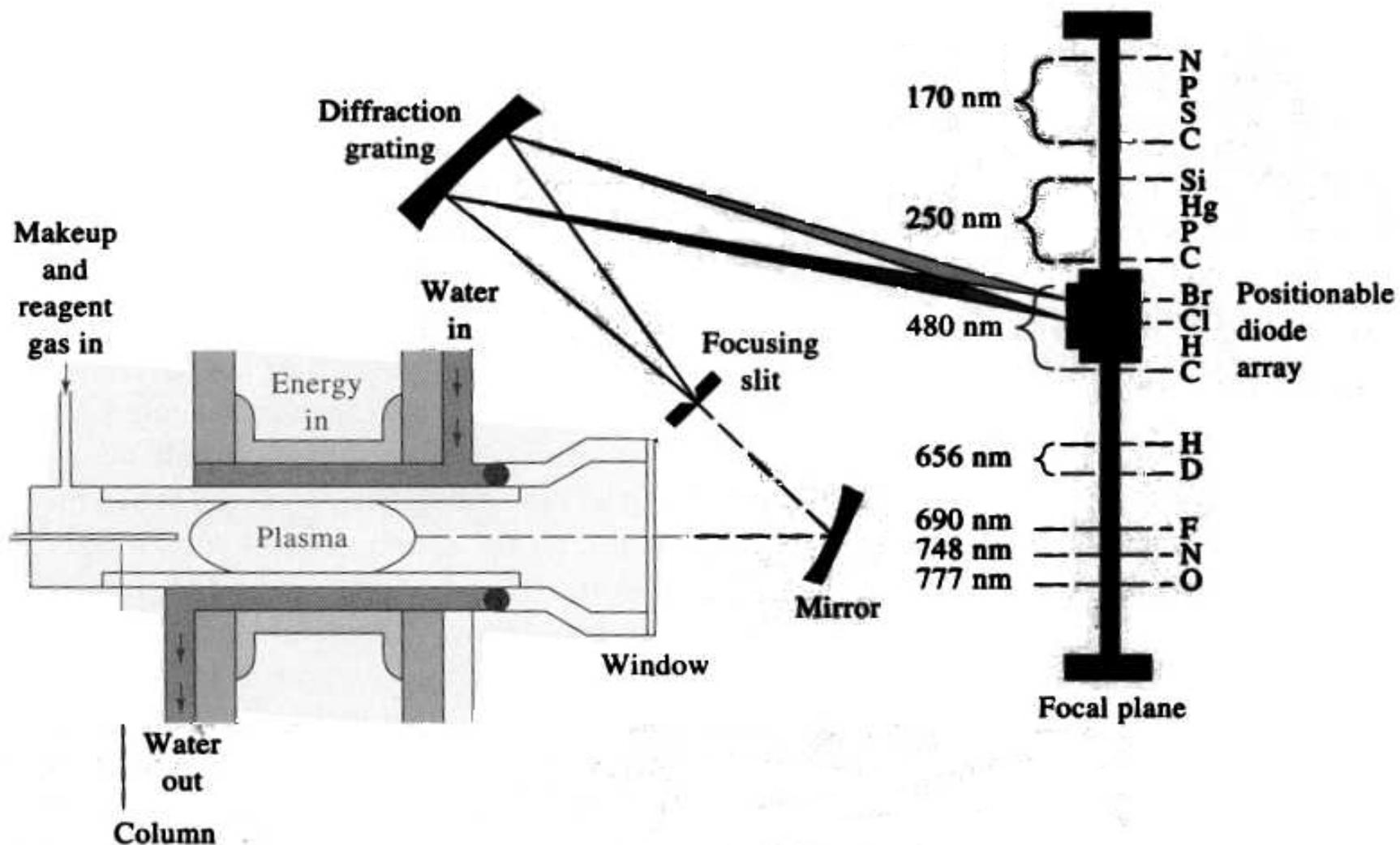
- Column effluent irradiated with intense UV light source
- Ionizes molecules
- Measure ions with electrodes in detector cell

# Unconventional Detectors (Hyphenated Techniques)

## Atomic Emission Detector (AED)

- Very powerful
- Sample eluent introduced to He microwave plasma atomizing all atoms in sample
- Uses diode array detector measuring optical emission over wide spectral range (170 - 780 nm)
- Measure many elements simultaneously

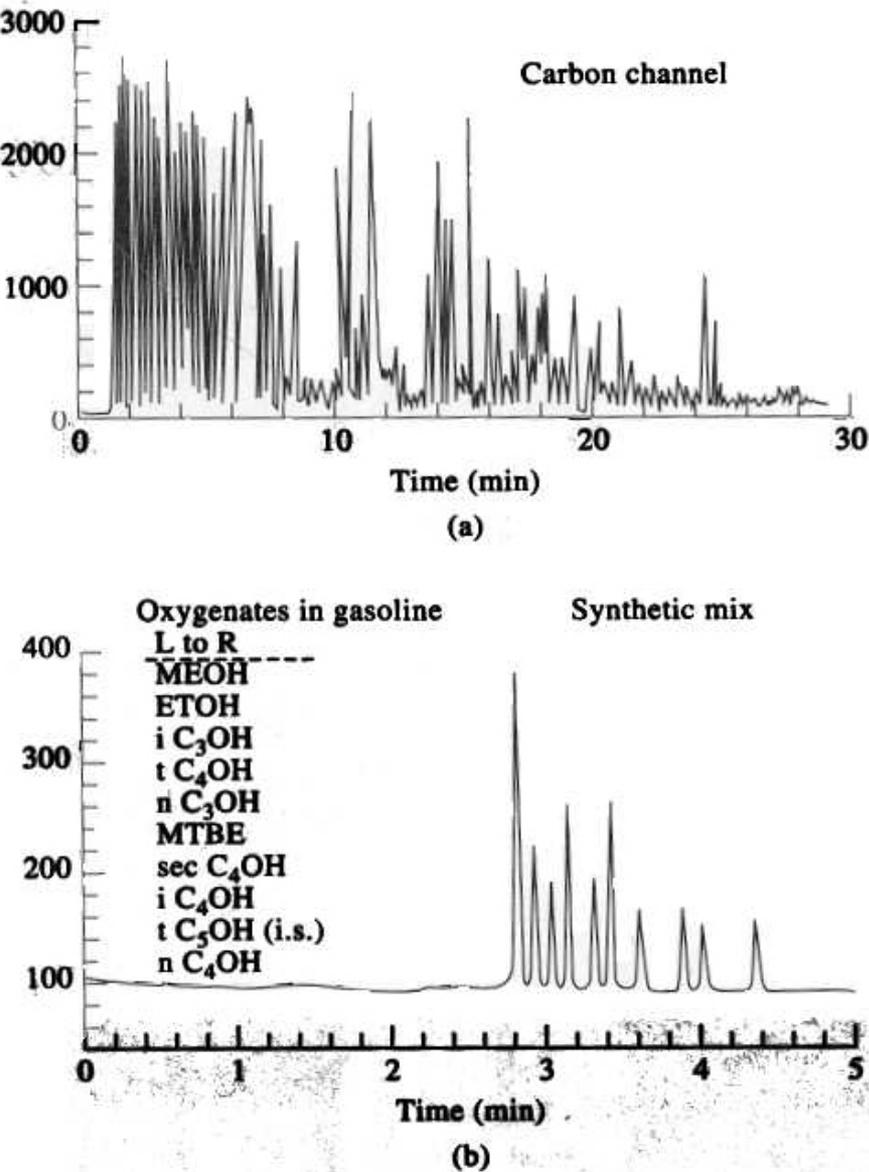
# GC-AED



**Figure 27-9** An atomic emission detector. (Courtesy of Hewlett-Packard Company.)

# GC-AED

- Potentially can measure 70 or more elements
- If look at C signal from AED get chromatogram with hundreds of peaks
- If look at O signal get very simple chromatogram with only a few peaks



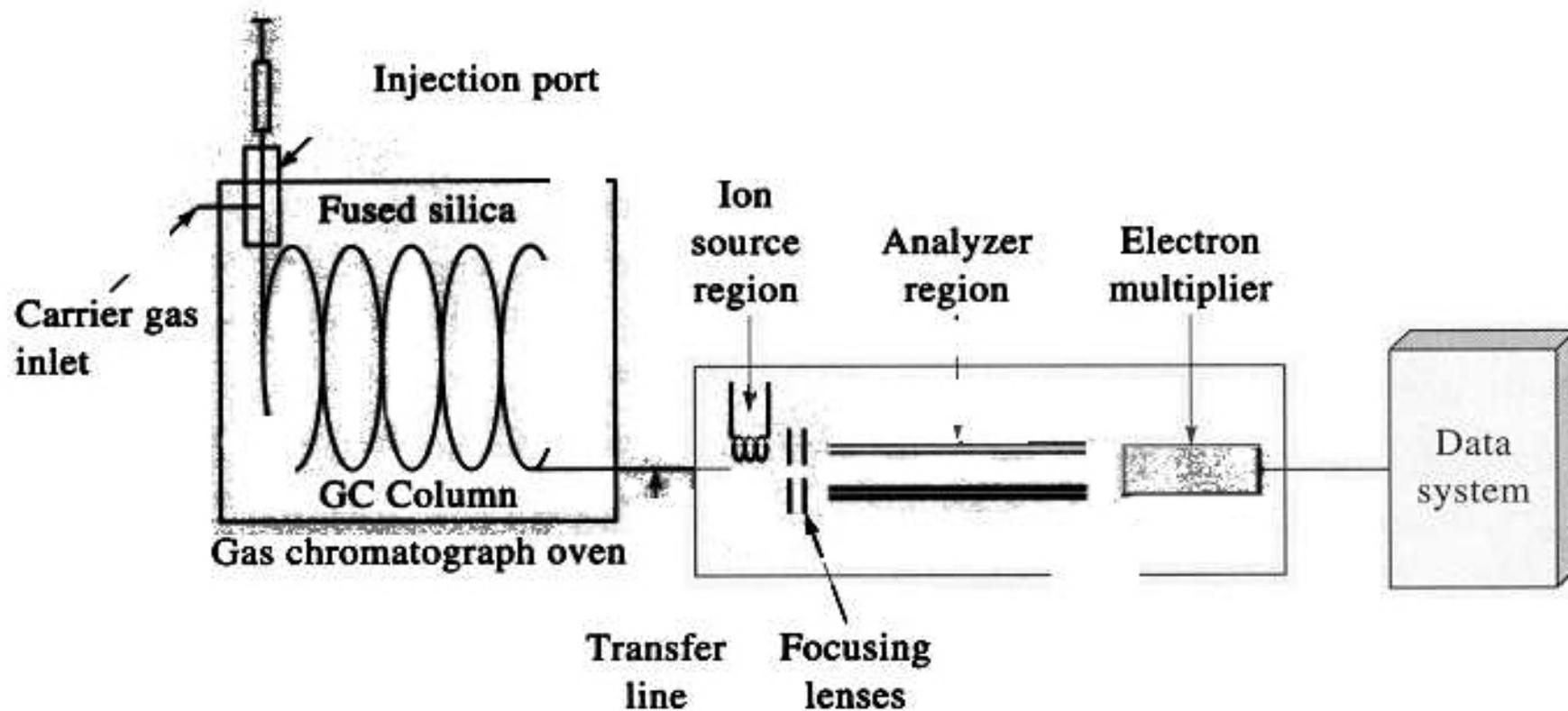
**Figure 27-10** Chromatograms for a gasoline sample containing a small amount of MTBE and several aliphatic alcohols: (a) monitoring the line for carbon; (b) monitoring the line for oxygen. (Courtesy of Hewlett-Packard Company.)

## GC – Mass Spectrometry (GC-MS)

- Interfacing GC & MS normally difficult
- GC at pressure above atmospheric while MS under high vacuum
- Need special interfaces for packed columns
  - Jet separator – discussed below
  - Membrane separator – a membrane sandwich between spiral channels, column effluent on one side under pressure, MS on other side under vacuum – relies on differential permeability of carrier gas vs analyte molecules

# GC-MS Schematic

Interface less critical for capillary columns



**Figure 27-13** Schematic of a typical capillary gas chromatography/mass spectrometer.

## Several types of Mass Specs available

- Rarely magnetic sector, time of flight (TOF) or ion cyclotron
- Usually quadrupole or ion trap for GC-MS
- Less expensive
- Less maintenance
- Easy to use
- Normally use electron multiplier as detector
- All MS systems need ion source, either electron impact or chemical ionization

## Three modes of operation for GC-MS

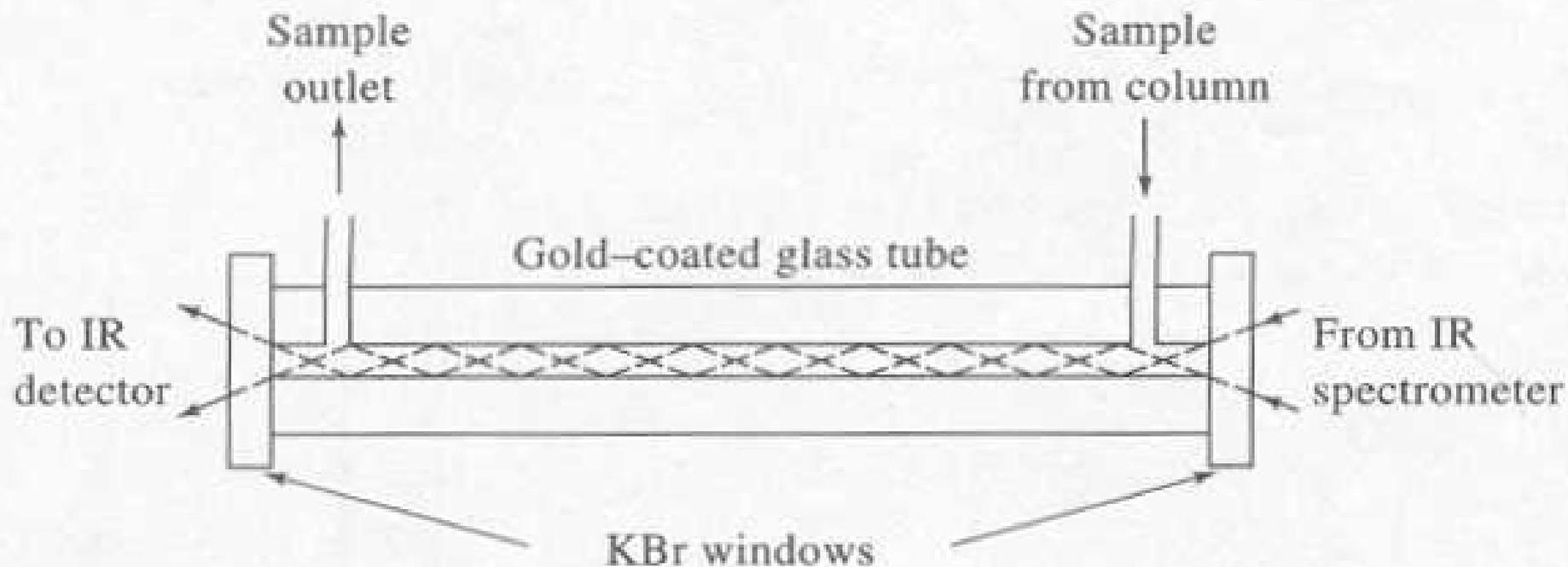
- 1) Spectral mode – look at mass spectrum every second or so during chromatogram - gives most information for research or method development
- 2) Total ion current – sum signal for all ions as one large signal – highest sensitivity
- 3) Selective ion monitoring (SIM) – look at certain mass/charge ratios for compounds of interest – routine analysis

# GC-MS

- sensitive
- can be very selective in SIM mode
- powerful for qualitatively & quantitatively
- used for complete unknowns
- used for court cases/expert witness

# GC-FTIR

- Powerful technique for identifying compounds
- Use heated light pipe 1 to 3 mm dia



**Figure 27-17** A typical light pipe for GC/IR instruments.

# GC-FTIR

- Powerful technique for identifying compounds
- Use heated light pipe 1 to 3 mm dia and 10 to 40 cm long
- Heat to prevent condensation of sample
- Cool detector for sensitivity
- Gives structural information from spectrum
- Not very common

# GC Columns & Stationary Phases

- Historically used packed columns
- Stationary phase coated as a thin film on a high surface area solid support
- Theoretical studies showed that unpacked columns with narrow diameters were better
- Open tubular columns first developed
- Capillary columns came later because
  - Very fragile, difficult to construct, hard to connect to GCs, small samples hard to detect, difficult to coat column walls, etc.

## Packed Columns

- Tubing of metal, glass, Teflon, etc.
- 2 to 3 m long and 2 to 4 mm in dia
- Packed with diatomaceous earth ( $\text{SiO}_2$ ), clay, carbon particles, glass microbeads, polymer
- Diameter 150-250  $\mu\text{m}$  (60-100 mesh) 1  $\text{m}^2/\text{g}$
- Thin coating of liquid stationary phase

**TABLE 27-2** Some Common Stationary Phases for Gas-Liquid Chromatography

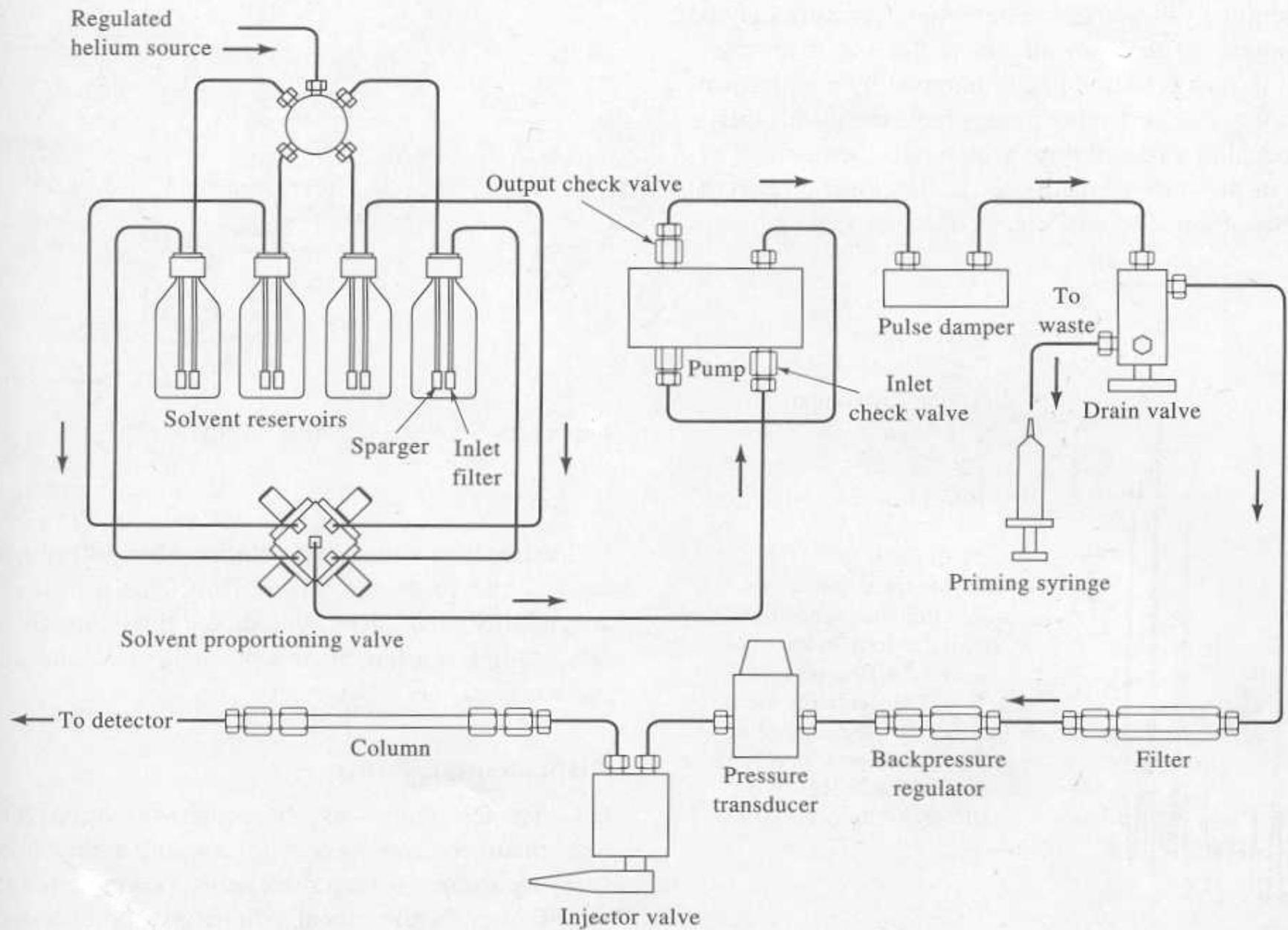
| Stationary Phase                                 | Common Trade Name | Maximum Temperature, °C | Common Applications  |
|--|-------------------|-------------------------|--|
| Polydimethyl siloxane                            | OV-1, SE-30       | 350                     | General-purpose nonpolar phase; hydrocarbons; polynuclear aromatics; drugs; steroids; PCBs |
| Poly(phenylmethyldimethyl) siloxane (10% phenyl) | OV-3, SE-52       | 350                     | Fatty acid methyl esters; alkaloids; drugs; halogenated compounds                          |
| Poly(phenylmethyl) siloxane (50% phenyl)         | OV-17             | 250                     | Drugs; steroids; pesticides; glycols   |
| Poly(trifluoropropyldimethyl) siloxane           | OV-210            | 200                     | Chlorinated aromatics; nitroaromatics; alkyl-substituted benzenes                          |
| Polyethylene glycol                              | Carbowax 20M      | 250                     | Free acids; alcohols; ethers; essential oils; glycols                                      |
| Poly(dicyanoallyldimethyl) siloxane              | OV-275            | 240                     | Polyunsaturated fatty acids; rosin acids; free acids; alcohols                             |

# High-Performance Liquid Chromatography (HPLC)

- Scope
- Instrumentation – eluants, injectors, columns
- Modes of HPLC
  - Partition chromatography
  - Adsorption chromatography
  - Ion chromatography
  - Size exclusion chromatography

# HPLC

- Most widely used separation technique
- Broad applicability – organic & inorganic
- Can be very sensitive, accurate & precise
- Suitable for separation of nonvolatile species
- Has found numerous uses in industry, clinical settings, environmental areas, pharmaceuticals, etc.



**Figure 28-4** Schematic of an apparatus for HPLC. (Courtesy of Perkin-Elmer Corporation, Norwalk, CT.)

Solvents (mobile phase) – are stored in special reservoirs connected to the pumping system – must be free of particles that can clog components & free of bubble forming gases that get trapped in column or detector

Three basic ways to degas solvents

- 1) vacuum or suction filter (0.4 or 0.2  $\mu\text{m}$ )
- 2) ultrasonicate (with vacuum)
- 3) He purge (sparge units often built in)

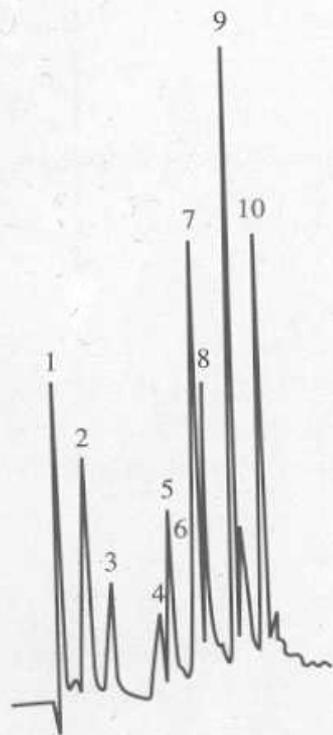
Can purchase HPLC solvents & water - still

In GC the analyte affinity for the column is influenced by temp

In HPLC the solvent strength affects an analytes retention on column

Therefore, analogous to temp programming in GC, do solvent programming in HPLC

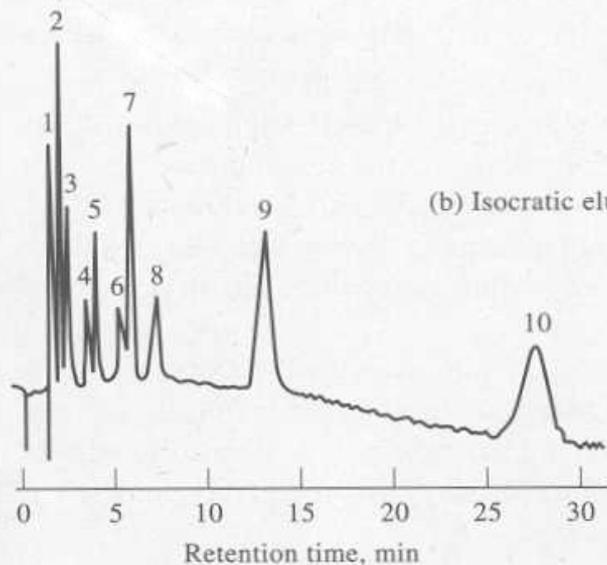
This is also referred to as gradient elution



(a) Gradient elution

Peak identity

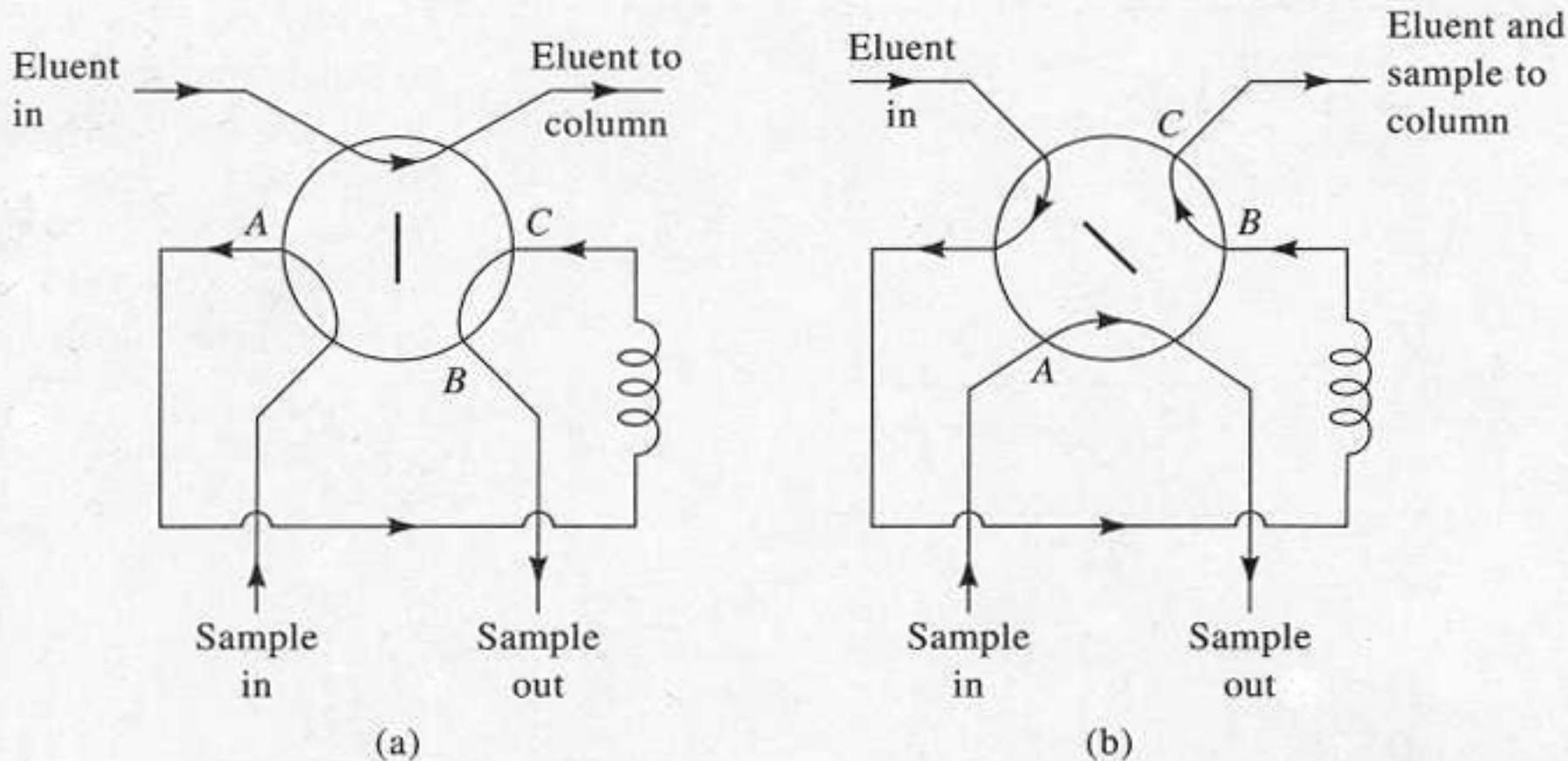
1. Benzene
2. Monochlorobenzene
3. Orthodichlorobenzene
4. 1,2,3-trichlorobenzene
5. 1,3,5-trichlorobenzene
6. 1,2,4-trichlorobenzene
7. 1,2,3,4-tetrachlorobenzene
8. 1,2,4,5-tetrachlorobenzene
9. Pentachlorobenzene
10. Hexachlorobenzene



(b) Isocratic elution

Gradient elution  
dramatically  
improves the  
efficiency of  
separation

HPLC sample injectors are 6 port rotary valves that are overfilled by syringe giving extreme accuracy & precision – typical volumes are 10 to 50  $\mu\text{L}$  but can be larger

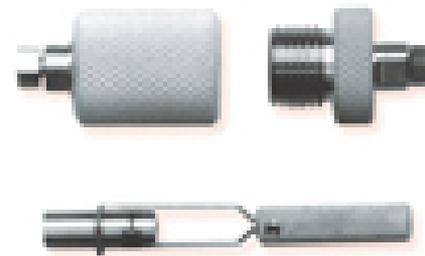
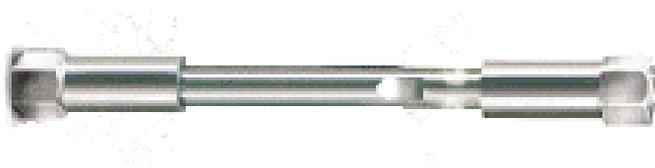


**Figure 27-4** A rotary sample valve: valve position (a) for filling sample loop A-C-B and (b) for introduction of sample into column.

# Columns

- usually stainless steel
- can be PEEK (poly ether ether ketone)
- may cost \$300-\$1000 packed
- Length 10-30 cm, ID 4-10 mm
- Packings are 3, 5, or 10  $\mu\text{m}$  particle size
- Most common 25 cm, 5  $\mu$ , 4.6 mm ID
- $N = 40,000$  to 60,000
- Normally packed under 6000 psi pressure at factory as a slurry

Guard columns are normally used before the analytical column to protect & increase lifetime of column – operator usually slurry or dry packs short guard column regularly with same or similar packing used in analytical column (old column material) – can purchase guard systems, cartridges, etc.



## Detectors for HPLC

- Ideal characteristics same as GC
- Exception is temp range
- Low dead volume 1 to 10  $\mu\text{L}$

Most common detector is **UV-vis absorbance**

Three types

- 1) Filter instrument – optical filters, Hg lamp
- 2) Variable wavelength – monochromator
- 3) Diode array detector- provide spectra

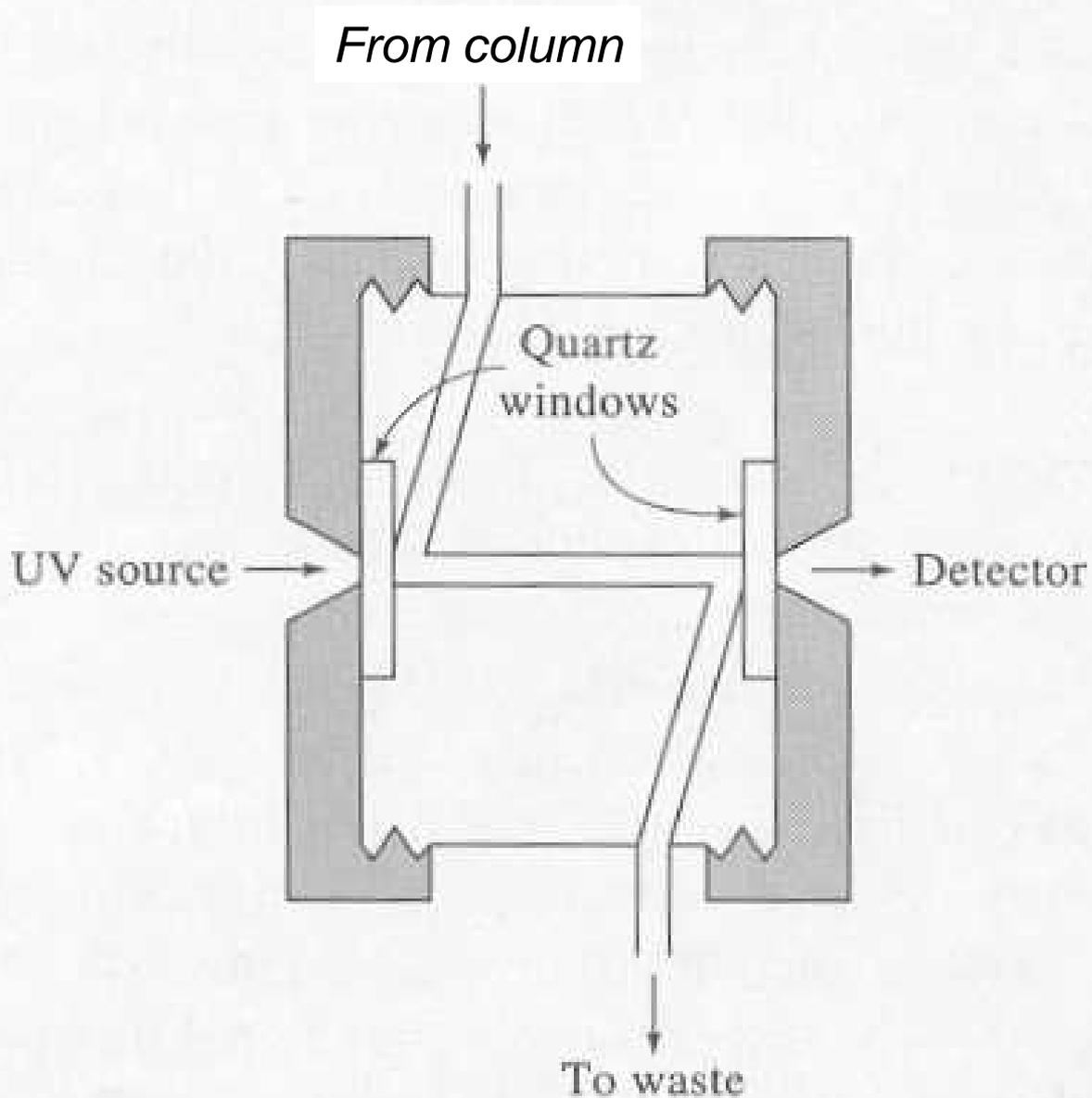
# Many HPLC detectors available

## For universal & selective detection

**TABLE 28-1 Performances of LC Detectors**

| LC Detector                   | Commercially Available | Mass LOD (commercial detectors) <sup>a</sup> | Mass LOD (state of the art) <sup>b</sup> |
|-------------------------------|------------------------|--|--|
| Absorbance                    | Yes <sup>c</sup>       | 100 pg–1 ng                                  | 1 pg                                     |
| Fluorescence                  | Yes <sup>c</sup>       | 1–10 pg                                      | 10 fg                                    |
| Electrochemical               | Yes <sup>c</sup>       | 10 pg–1 ng                                   | 100 fg                                   |
| Refractive index              | Yes                    | 100 ng–1 $\mu$ g                             | 10 ng                                    |
| Conductivity                  | Yes                    | 500 pg–1 ng                                  | 500 pg                                   |
| Mass spectrometry             | Yes <sup>d</sup>       | 100 pg–1 ng                                  | 1 pg                                     |
| FT-IR                         | Yes <sup>d</sup>       | 1 $\mu$ g                                    | 100 ng                                   |
| Light scattering <sup>e</sup> | Yes                    | 10 $\mu$ g                                   | 500 ng                                   |
| Optical activity              | No                     | —  | 1 ng                                     |
| Element selective             | No                     | —  | 10 ng                                    |
| Photoionization               | No                     | —  | 1 pg–1 ng                                |

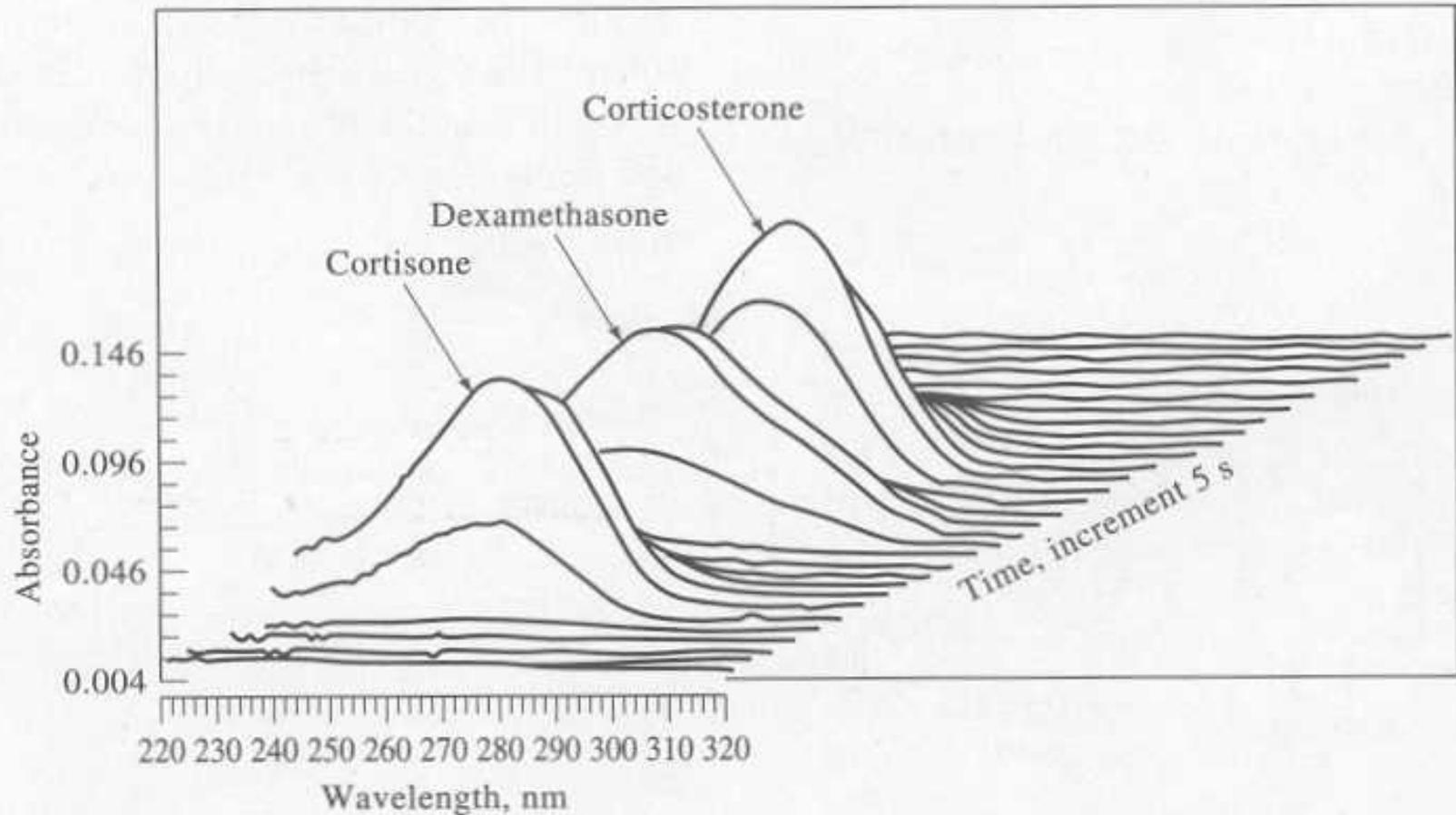
- 1) Filter based UV-vis detector – Typically set at 254 nm using the most prominent band in Hg spectrum – can also use 313, 365, 334 nm and other lines as well
- 2) Variable wavelength detectors – use continuum source like ( $D_2$  or  $H_2$ ) & a monochromator, select any  $\lambda$ , less sensitive
- 3) PDA -  $D_2$  or  $H_2$  source, disperse & focus on diode array, get complete spectrum every 1 sec, powerful, expensive, less sensitive, lots of data generated



Cell for  
UV-vis  
detector  
for HPLC  
- Low vol

**Figure 28-9** Ultraviolet detector cell for HPLC.

# Diode Array Detector



**Figure 28-10** Absorption spectra of the eluent from a mixture of three steroids taken at 5-second intervals. (Courtesy of Hewlett-Packard Company, Palo Alto, CA.)

Fluorescence detector – normally  
fixed wavelength filter fluorometer  
excitation filter & emission filter  
can be changed for particular  $\lambda$  of interest  
gives selectivity based on:

- ability to exhibit fluorescence
- excitation wavelength
- emission wavelength

Variable  $\lambda$  monochromator based  
fluorescence detectors also available

Filter based detectors usually more sensitive

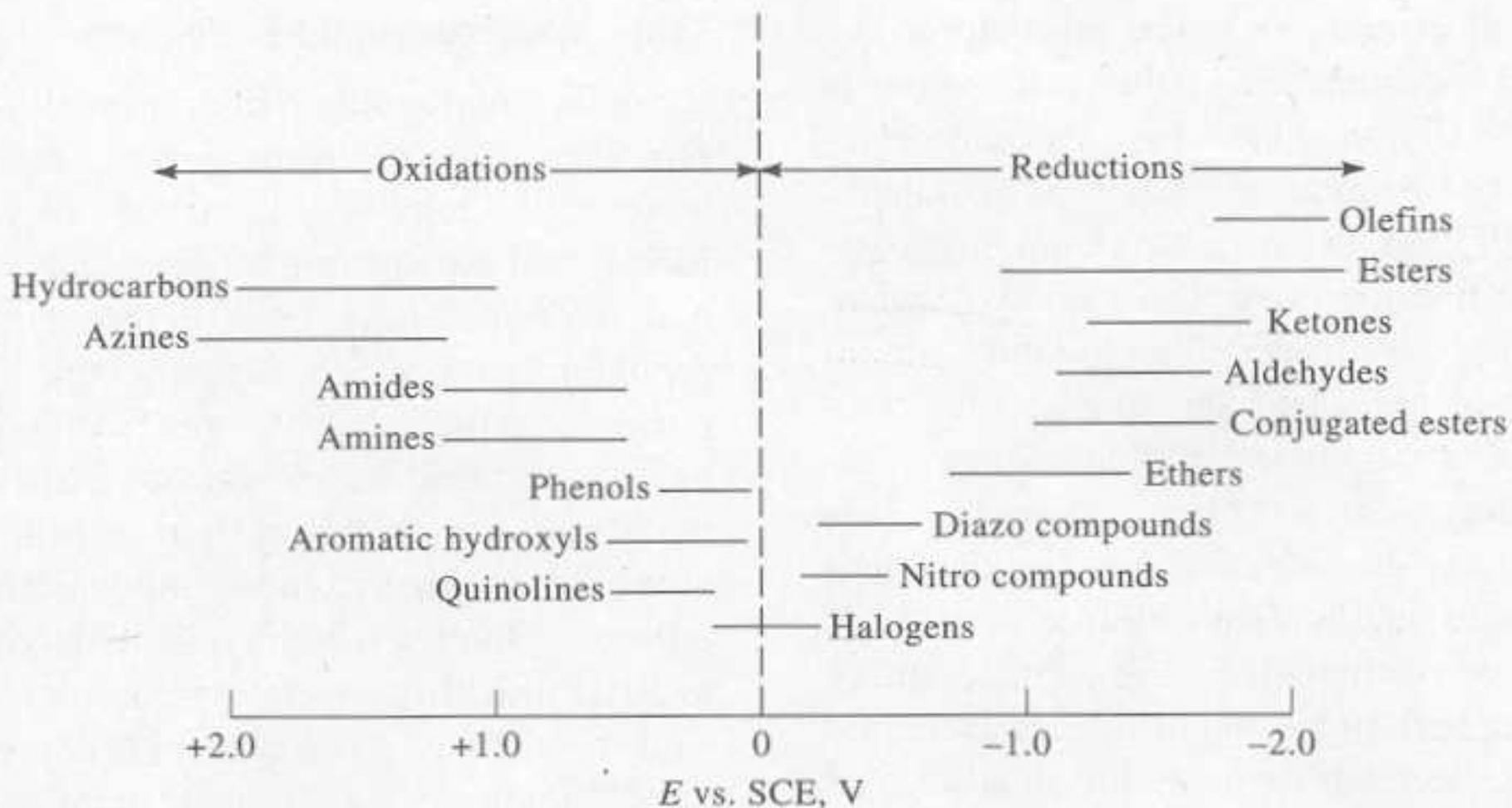
Refractive index detector (RI) -  
responds to nearly all solutes  
but has poor sensitivity – detects  
changes in refractive index as sample  
passes through as long as solute has  
different RI than solvent – analogous to  
TCD in GC

# Electrochemical Detection

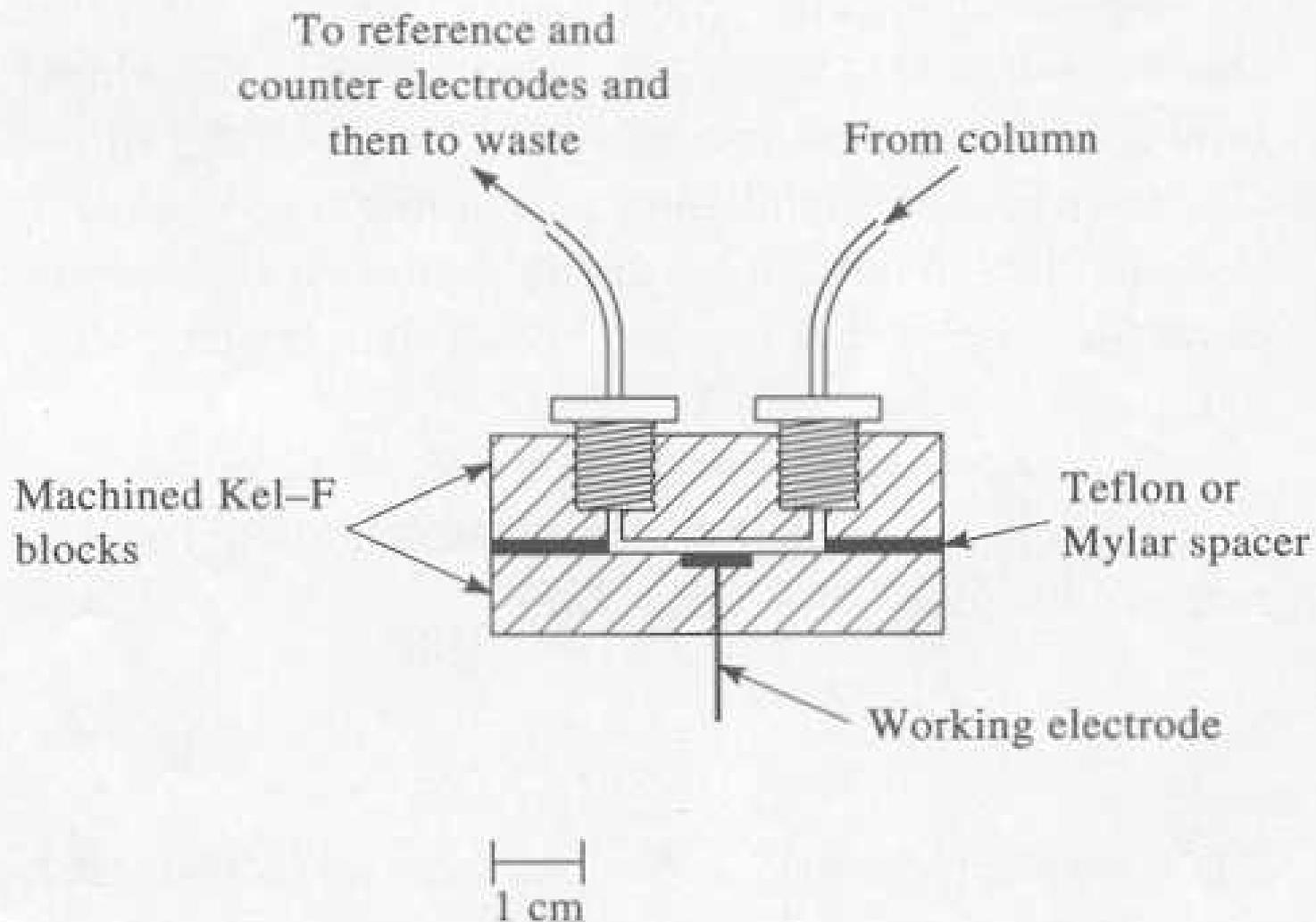
- Amperometric – fix potential & measure current ( $i$ )
- Conductometric – measure conductivity
- Coulometric – fix potential & integrate  $i$
- Voltammetric – vary potential & measure  $i$
- Potentiometric – measure potential

Can use 2 or 3 electrode design with Pt or carbon electrodes (glassy C or C paste)

Electrochem. detector nearly universal



**Figure 28-12** Potentially detectable organic functional groups by amperometric measurements. The horizontal lines show the range of oxidation or reduction potentials wherein compounds containing the indicated functional groups are electroactive.



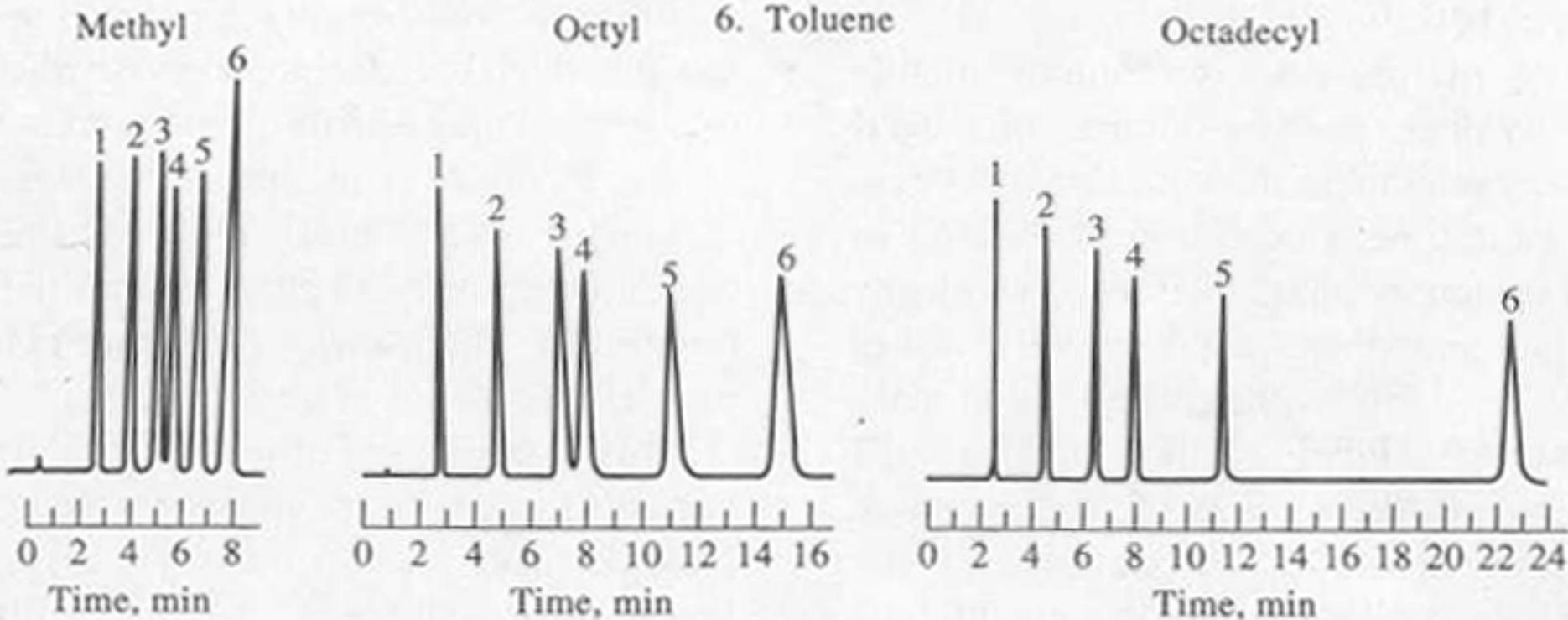
**Figure 28-13** Amperometric thin-layer detector cell for HPLC.

## Other HPLC detectors

- LC-MS using thermospray – new popularity (pharmaceuticals)
- Evaporative light scattering - polymers
- LC-FTIR
- LC-plasma emission or ICP-MS

Peak identification

1. Uracil
2. Phenol
3. Acetophenone
4. Nitrobenzene
5. Methyl benzoate
6. Toluene



**Figure 28-15** Effect of chain length on performance of reversed-phase siloxane columns packed with 5- $\mu\text{m}$  particles. Mobile phase: 50/50 methanol/water. Flow rate: 1.0 mL/min.

Besides  $C_{18}$  can have  $C_8$ ,  $C_4$ ,  $C_3$ ,  
 $C_2$ ,  $C_1$  plus functionalities like  
cyano ( $-C_2H_4CN$ ), amino ( $-C_2H_4NH_2$ ),  
diol ( $-C_3H_6O-CH_2-CHOHCH_2OH$ )

Each has different polarity

Can also do Ion Pair Chromatography or  
Paired-Ion Chromatography – type of RP-  
HPLC used to separate ionic species

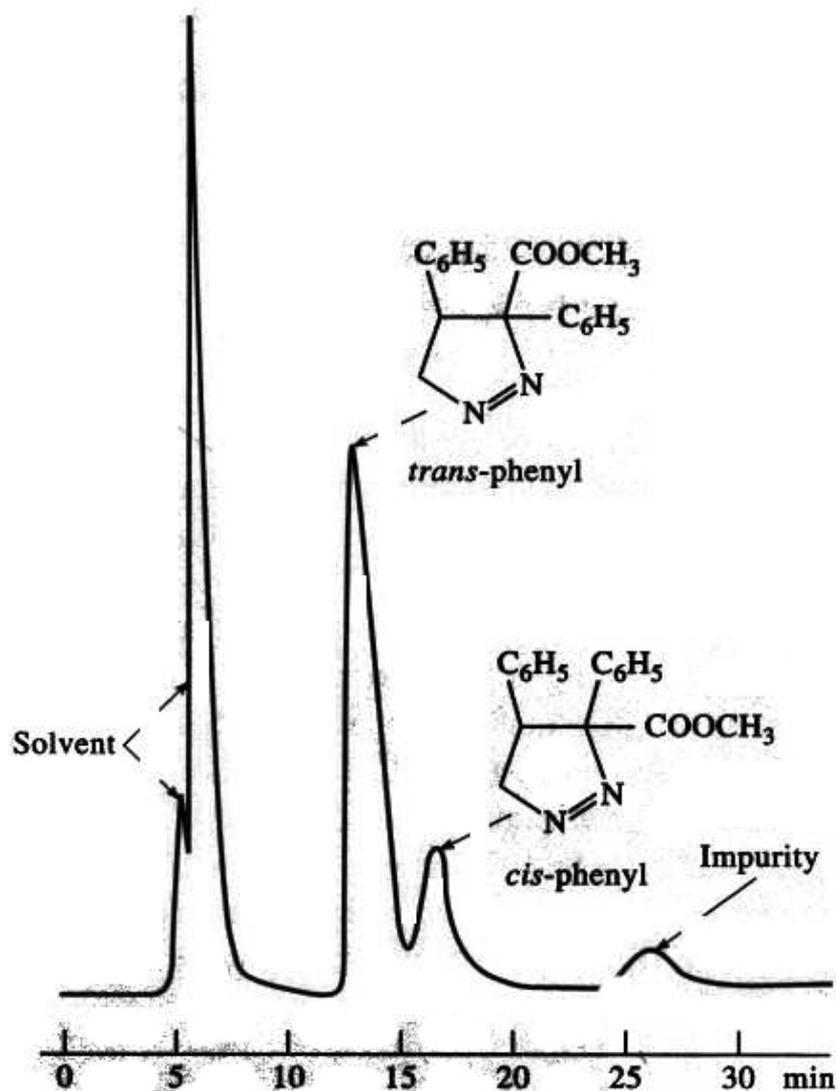
Still partition chrom. but use a reagent like a  
quaternary ammonium salt  $(C_4H_9)_4N^+$  to  
pair with analyte ions to separate by RP

# Adsorption Chromatography –

bare silica or alumina to separate non-polar compounds because they adsorb to the stationary phase & are eluted by adjusting solvent strength of mobile phase – important non-linear applications

Adsorption chrom. = normal phase chromatog.

Least popular mode of separation due to:  
strong adsorption, surface changes over time, with pH or water content



- Sample of an application of adsorption chromatography

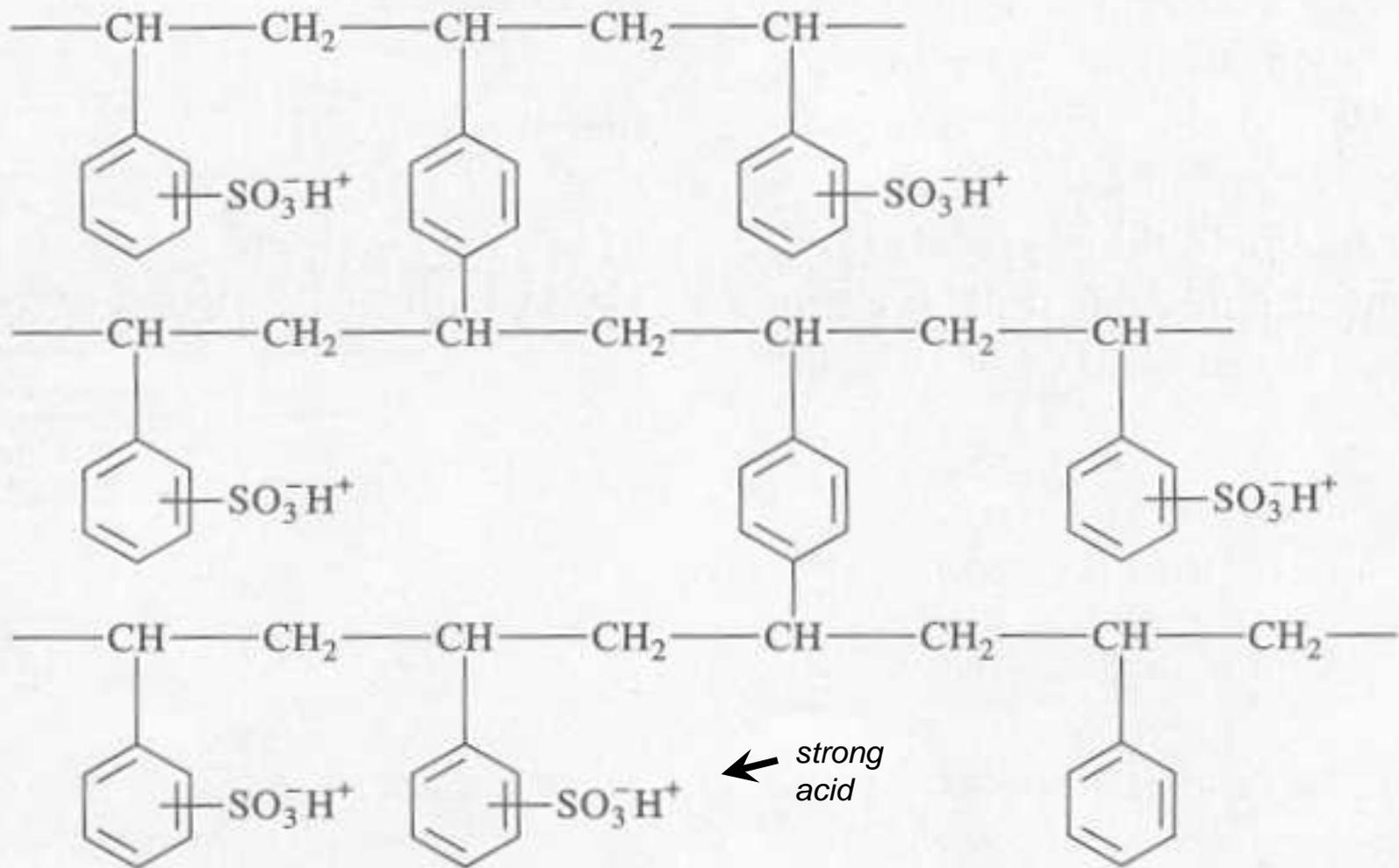
**Figure 28-20** A typical application of adsorption chromatography: separations of *cis*- and *trans*-pyrazoline. Column: 100 × 0.3 cm pellicular silica. Mobile phase: 50% methylene chloride/isooctane. Temperature: ambient. Flow rate: 0.225 mL/min. Detector: UV, 254 nm.

## Ion Chromatography (Ion Exchange)

Historically was developed for the Manhattan Project (atomic bomb)

Generally not automated because of the lack of good detectors until it was reinvented in 1970's at Dow Chemical using conductivity detection & chemical suppression

Stationary phases are resin beads of styrene-divinylbenzene functionalized with cationic & anionic groups developed for water purification in 1930's



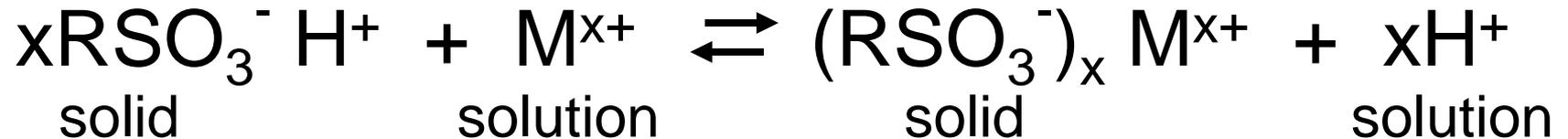
**Figure 28-21** Structure of a cross-linked polystyrene ion-exchange resin. Similar resins are used in which the —SO<sub>3</sub><sup>-</sup>H<sup>+</sup> group is replaced by —COO<sup>-</sup>H<sup>+</sup>, —NH<sub>3</sub><sup>+</sup>OH<sup>-</sup>, and —N(CH<sub>3</sub>)<sub>3</sub><sup>+</sup>OH<sup>-</sup> groups.

↑  
weak base

↑  
strong base

↑  
weak acid

Can write reactions in general format



Where R = polymer support (styrene divinylbenzene)

Can write equilibrium expression for exchange

$$K_{\text{ex}} = \frac{[(\text{RSO}_3^-)_x \text{M}^{x+}]_s [\text{H}^+]_{\text{aq}}^x}{[\text{RSO}_3^- \text{H}^+]_s^x [\text{M}^{x+}]_{\text{aq}}^x}$$

tells affinity of  
resin for  $\text{M}^+$   
compare to  $\text{H}^+$   
here or any ion

# Ion Exchange Process

Analyte ions ( $M^{x+}$ ) are passed thru column & retained on an ion-exchange site. The mobile phase contains some  $H^+$  & this is increased sufficiently to cause exchange with  $M^{x+}$ .

Back to equilibrium expression

$$K_{\text{ex}} = \frac{[(\text{RSO}_3^-)_x \text{M}^{x+}]_s [\text{H}^+]_{\text{aq}}^x}{[\text{RSO}_3^- \text{H}^+]_s [\text{M}^{x+}]_{\text{aq}}^x}$$

Rearrange to

$$\frac{[\text{RSO}_3^- \text{H}^+]_s}{[\text{H}^+]_{\text{aq}}^x} K_{\text{ex}} = \frac{[(\text{RSO}_3^-)_x \text{M}^{x+}]_s}{[\text{M}^{x+}]_{\text{aq}}^x}$$

During elution  $[\text{H}^+]$  is high &  $[\text{RSO}_3^- \text{H}^+]_s$  is high  
Left hand side of equation essentially constant

$$K = \frac{[(\text{RSO}_3^-)_x \text{M}^{x+}]_s}{[\text{M}^{x+}]_{\text{aq}}^x} = \frac{C_s}{C_M}$$

K turns out to be a distribution ratio (partition)

Order of affinity for sulfonated cation exchange

$\text{Tl}^+ > \text{Ag}^+ > \text{Cs}^+ > \text{Rb}^+ > \text{K}^+ > \text{NH}_4^+ > \text{Na}^+ > \text{H}^+ > \text{Li}^+$

$\text{Ba}^{2+} > \text{Pb}^{2+} > \text{Sr}^{2+} > \text{Ca}^{2+} > \text{Ni}^{2+} > \text{Cd}^{2+} > \text{Cu}^{2+} > \text{Co}^{2+} >$

$\text{Zn}^{2+} > \text{Hg}^{2+}$

# Ion Chromatography Detection

Basic detector is conductivity, but others are used such as UV-vis & atomic spectrometry (AA, AE) for metals

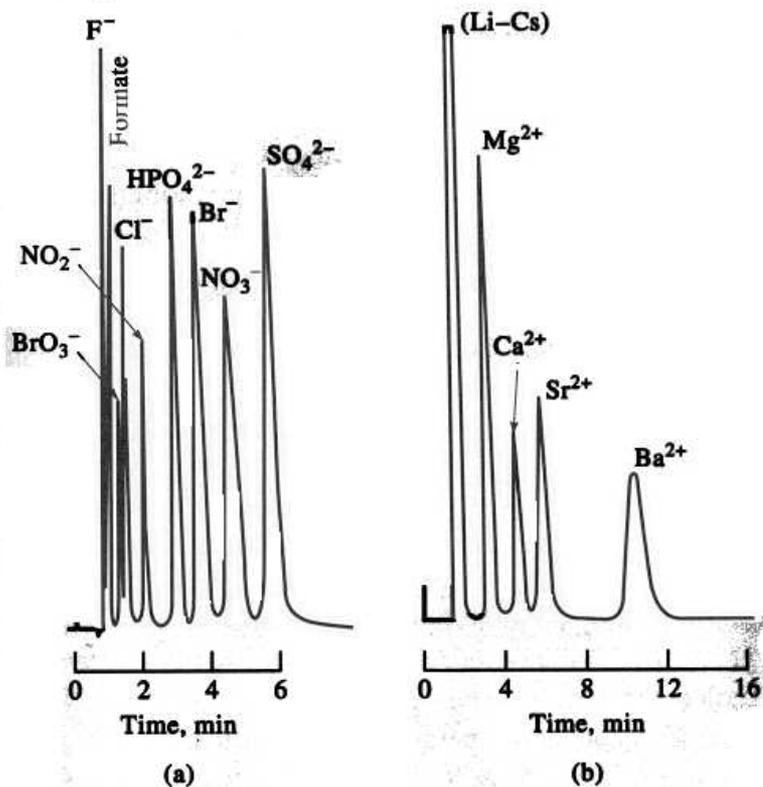
Measure conductivity change in effluent when analyte passes through

Problem – use high  $[H^+]$  to elute small  $[M^{x+}]$  which makes it difficult to detect  $[M^{x+}]$  conductivity on high background of  $[H^+]$

This problem hindered development of IC until the innovations made at Dow in 70's

| Concentrations, ppm            |    |
|--------------------------------|----|
| F <sup>-</sup>                 | 3  |
| Formate                        | 8  |
| BrO <sub>3</sub> <sup>-</sup>  | 10 |
| Cl <sup>-</sup>                | 4  |
| NO <sub>2</sub> <sup>-</sup>   | 10 |
| HPO <sub>4</sub> <sup>2-</sup> | 30 |
| Br <sup>-</sup>                | 30 |
| NO <sub>3</sub> <sup>-</sup>   | 30 |
| SO <sub>4</sub> <sup>2-</sup>  | 25 |

| Concentrations, ppm |    |
|---------------------|----|
| Ca <sup>2+</sup>    | 3  |
| Mg <sup>2+</sup>    | 3  |
| Sr <sup>2+</sup>    | 10 |
| Ba <sup>2+</sup>    | 25 |



**Figure 28-23** Typical applications of ion chromatography. (a) Separation of anions on an anion-exchange column. Eluent: 0.0028 M NaHCO<sub>3</sub>/0.0023 M Na<sub>2</sub>CO<sub>3</sub>. Sample size: 50 μL. (b) Separation of alkaline earth ions on a cation-exchange column. Eluent: 0.025 M phenylenediamine dihydrochloride/0.0025 M HCl. Sample size: 100 μL.

(Courtesy of Dionex Corporation, Sunnyvale, CA.)

- Typical IC separations

# Size Exclusion Chrom. (SEC)

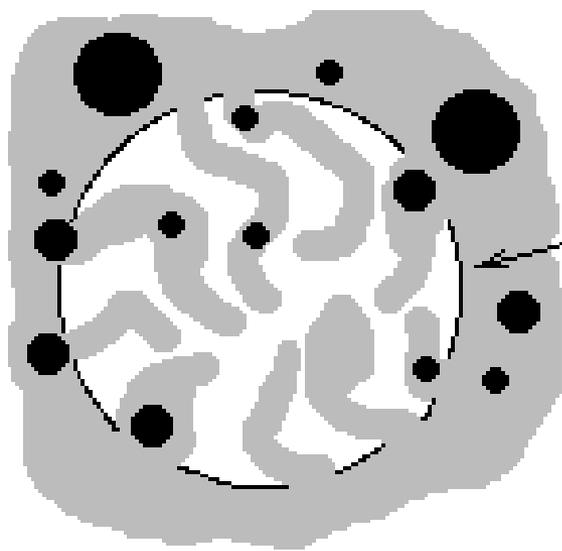
Packings are porous polymeric (resins) or silica based materials

Two names used for the same process:

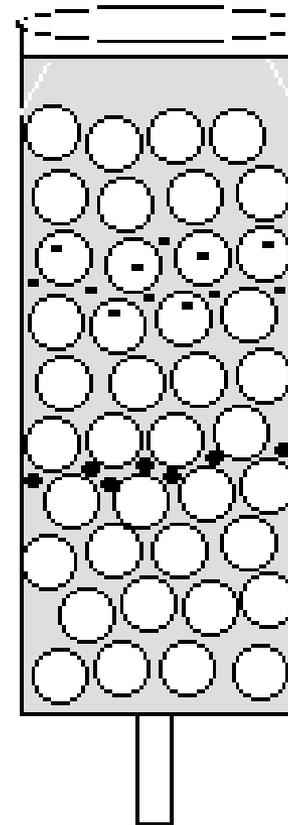
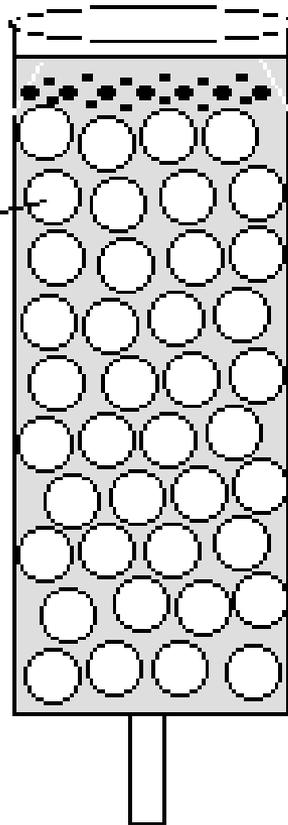
- 1) Gel filtration chrom. = aqueous solvent
- 2) Gel permeation chromatography = non-aqueous mobile phase

Column packing works like a molecular filter allowing small molecules access to every pore, retarding their progress – large molecules pass thru more quickly

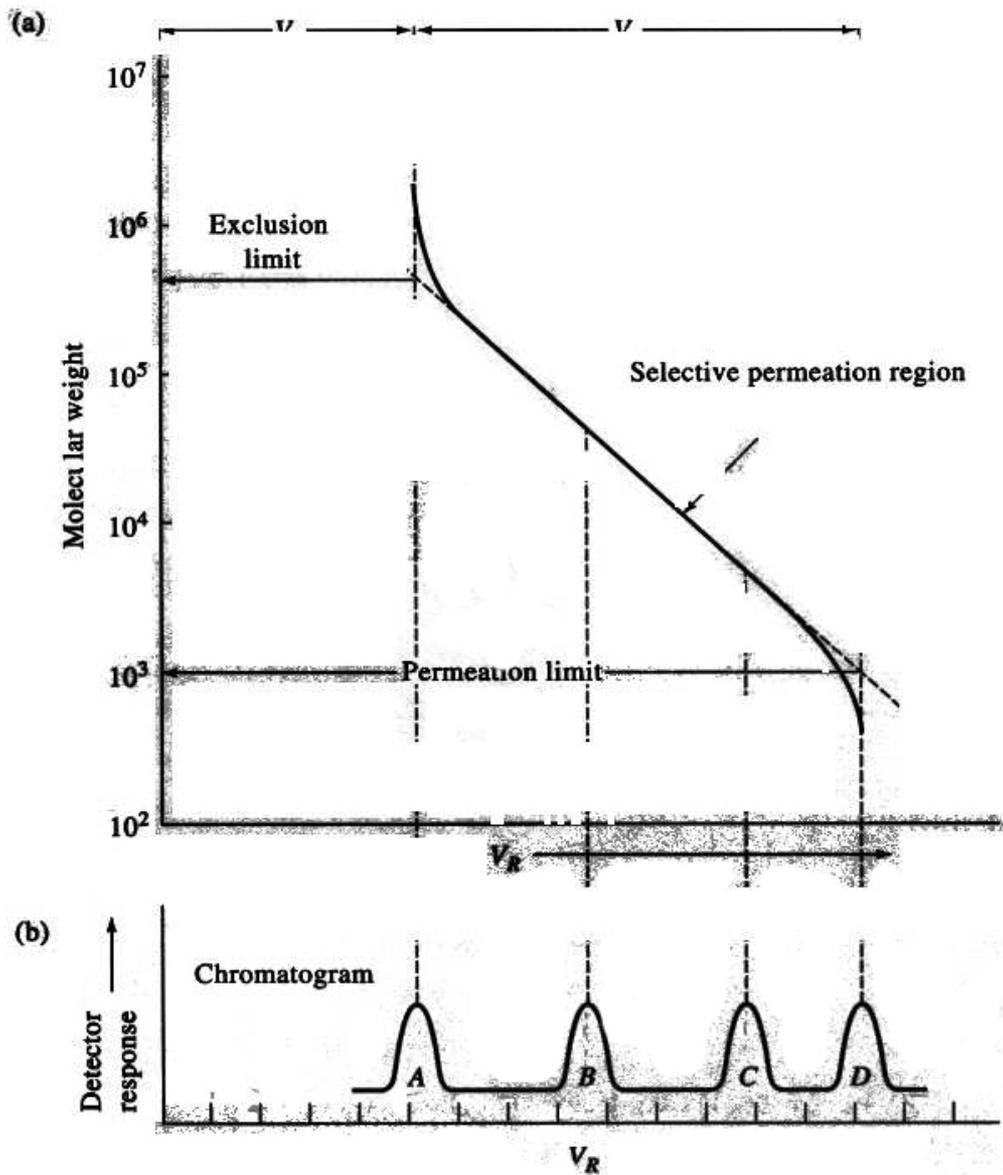
# SEC



Gel beads have pores in them of a defined size range which allows smaller molecules to enter but excludes molecules larger than the pore diameters.



- ← Molecules smaller than gel bead pores
- ← Molecules larger than gel bead pores



**Figure 28-27** (a) Calibration curve for a size-exclusion column. (b) Chromatogram showing peak A containing all compounds with molecular weights greater than the exclusion limit, peaks B and C consisting of compounds within the selective permeation region, and peak D containing all compounds smaller than the permeation limit.